

ECOLOGICAL STATE OF LAKE DUROWSKIE ALGAE AS AN ASSESSMENT INDICATOR

26th June – 30th June 2017



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Introduction

Wagrowiec is a town situated northwest of Poland. This town is known for tourism destination because this city has a beautiful lake - Lake Durowskie. Tourism has been one of the major incomes generating sector for the city. In addition, people also highly rely on the lake for water consumption, irrigation, and other water-based activities. Lake Durowskie is the last of a five chain lakes that a connected together.

Due to heavy use of the lake for recreational activities and the discharge of nutrient rich water from the upper lakes there was a sudden bloom of cyanobacteria in Lake Durowskie. Cyanobacteria in a high density can be harmful for lakes as they produce toxins that are harmful for human and aquatic wildlife. As a result cyanobacteria bloom the beach area along the lake was closed down because it was described as unhealthy for human use (fishing, swimming etc). Water quality deteriorated in time. Algal bloom is a typical characteristic of lakes, rivers and other water bodies that have been heavily polluted. Bloom may lead to oxygen depletion, fish gill clogging and the production of bad odours (Svirčev et al. 2014).

Due the social and economic importance of Lake Durowskie to the people of Durowskie there was a need to device restoration and subsequent management of the lake. Mitigation actions were suggested to restore Lake Durowskie into a good quality state and in 2009 restoration of the lake begun. Since the commencement of restoration there has been the need to monitor the recovery of the lake. Monitoring has relied on the use of biological and chemical indicators to assess the ecological state of the lake. Biological indicators include the use of phytoplankton, macrophytes and macro-invertebrate communities. Several studies have suggested the importance of using phytoplankton as indicators of water quality (Bianchi et al. 2003). Due to their sensitivity to environmental conditions, phytoplankton presents as a useful species group to evaluate changes in water bodies and as early warning signals of water pollution (Tas et al. 2009). Phytoplankton is therefore used in this study to assess the ecological state of the lake in the course of eight years of restoration. Periphyton is also taken into account as an indicator of water quality. Periphyton occupy submerged surfaces and are usually found on stones and submerged plants. Since they have special habitat requirement they are particularly good indicators for polluted waters.

Aims and objectives

The main aim of this project is to assess the ecological state of Lake Durowskie using phytoplankton and periphyton as bio-indicators. Also a comparative analysis was done to assess the trend and progress of restoration.

Materials and Methods

Study area

The study was conducted in Lake Durowskiej. Lake Durowski lies with the coordinates N 52° 49' 06" and E 17° 12' 01" in the Polish city Wągrowiec and is the final lake in a chain of five lakes that are connected by rivers. The profile of Lake Durowskie are given below (Table 1).

Table 1: Profile of Lake Durowskie.

Morphometric parameter		Measurements
Surface Area (ha)		143.7
Volume (m ³)		11322900
Maximum depth (m)		14.6
Average depth (m)		7.9
Total catchment area (km ²)		236.1
Direct catchment area (ha)		1581.3
Land use of surrounding area	Agriculture (%)	58.26
	Forest (%)	33.52
	Urban (%)	8.25

Sampling design

Phytoplankton sampling was carried out at 8 sites in the lake while periphyton sampling was done in 12 sites along the shore (Figure 1). Water samples were taken from depths of 0 m, 1 m, 2 m and 3 m to analyse the vertical distribution of phytoplankton. Thirty litres of water was sampled from the

various depth using a water sampler. Water collected was then filtered with a plankton net. Filtrates collected (periphyton and phytoplankton) were fixed with Lugol's iodine in the field.

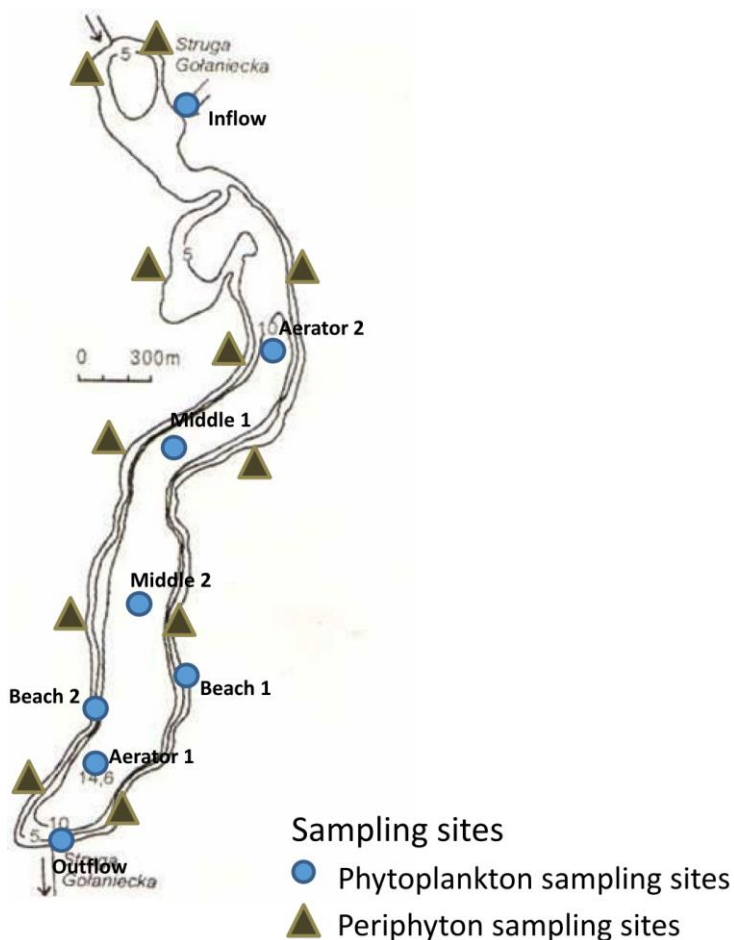


Figure 1: Phytoplankton and Periphyton sampling sites.

In addition to the phytochemical and periphyton samples taken, physico-chemical parameters were also measured (Table 2).

Table 2: Physico-chemical parameters measured in different sampling locations in Lake Durowskie.

Parameter	Unit
Temperature	°C
pH	
Conductivity	μScm^{-1}
Transparency	M
Conductivity	μScm^{-1}

Laboratory analysis

Phytoplankton and periphyton that were sampled were identified and counted as the number of individuals per litre of water (number/L)

The number of individuals for each species group was counted in 100 cells under the microscope. Conversion factor was determined using the following equation.

sample concentration 30 mL from 30 L; $30\ 000\ \text{mm}^3$: $1.25\ \text{mm}^3 = 24\ 000$

$24\ 000 - 30\ \text{L}$

$x - 1\ \text{L}$

$x = 800 - \text{factual concentration}$

Counts of individuals in 100 cells was multiplied by 800 to obtain the individual number of cells per liter.

Biomass Measurements (mg/L)

To estimate the biovolume of 1 cell of particular algal species,

- Multiplying biovolume of 1 cell with the cell count in 1mL
- Divide by 10^9 to make it under the unit of mgL^{-1}

The biomass was always given to 4 decimal places.

Data analysis

Data received from periphyton sampling was used to map the distribution of red algae and subsequently the calculation of Trophic Diatom Index. Data obtained from phytoplankton survey was used in calculating Mixed index, Jaccard index, Shannon-Weaver Diversity index and PMPL index. Background of each index is given blow.

Periphyton

Diatom Index

Diatom index is an informative tool used in determining the ecological state of lakes. This index requires that at least 10 species that are sensitive to trophic level should be present in a given sample. To calculate Diatom index, trophic index (TJ), the index of referential species (pGR) and their standardization are needed.

Trophy index (TJ)

$$TJ = \frac{(TJ_i \times wTJ_i \times Li)}{(wTJ_i \times Li)};$$

Where,

TJ_i - sensitivity of species for the trophic state;

wTJ_i - range of the tolerance of the algal species;

Li - number of specimens of the determined species divided by the number of all identified individuals in the sample

Index of referential species (pGR)

$$pGR = \frac{NB - (NC + ND)}{NB + NC + ND};$$

Where,

NB - number of referential species for all lakes;

NC - number of referential species for deep lakes and of degradation species in shallow lakes; ND - number of degradation species for both kinds of lakes

Transformation of standardized value in the range from 1 to 0,

$$Z - TJ = 1 - ((TJ - 1) \cdot 0.25)$$

$$Z - pGR = (pGR + 1) \cdot 0.5$$

Finally Diatom index (D) was calculated by the following equation,

$$DI = (Z - TJ + Z - pGR) / 2$$

The value obtained from Diatom index describes the ecological state of a lake as given below.

Diatom index value	Class
> 0,83	Very good
0,55 - 0,82	Good
0,30 - 0,54	Moderate
0,15 - 0,29	Poor
< 0,15	Bad

Phytoplankton

Mixed Index of Nygaard

This method depends on number of species from all different taxonomical groups.

$$\text{Mixed Index} = \frac{\text{Cyanobacteria} + \text{Chlorococcales} + \text{Centric diatoms} + \text{Euglenoids} + \text{Desmids}}{\text{Total species}}$$

The value obtained for mixed index implies different trophic level classes as described below.

Dystrophy	0.0 - 0.2
Oligotrophy	0.2 - 1.0
Mesotrophy	1.0 - 3.0
Eutrophy	3.0 – 5.0
Hypertrophy	5.0 – 43.0

Jaccard Index (Jaccard, 1912)

This index is used in comparing species across different study sites. Absence and presence data of species is used.

$$S_J = a / (a + b + c)$$

Where,

S_J = Jaccard similarity index;

a = number of species common to (shared by) site;

b = number of species unique to the first site;

c = number of species unique to the second site.

Diversity Index

Shannon-Wiener diversity and evenness indices

The diversity and evenness of phytoplankton species was measured across different sites. The formula for Shannon diversity and evenness is given below.

$$H' = -\sum_{i=1}^s p_i \log(p_i);$$

Where,

H' - Shannon index;

p_i – relative abundance of each species in the site

$$E = \frac{-\sum p_i \times \log(p)}{\log(S)};$$

E – Evenness (equitability);

S – Total number of species in each site

Phytoplankton Multimetric for Polish Lakes (PMPL Index)

The PMPL is an ecological state indicator that was used to implement the European Water Framework Directive for Lakes in Poland. Computing this index requires three parameter which include Chlorophyll a, Total biomass and Cyanobacteria biomass. All three single metrics and PMPL index value ranging from 1 to 5.

$$PMPL = \frac{[YCh + YBm + YCy]}{3}$$

Where,

YCh - Chlorophyll-a concentrations

YBm - General biomass of phytoplankton

YCy - Biomass of cyanobacteria

Value obtained for PMPL is indicating different classes of tropic levels as below:

Ecological status	PMPL
very good	0,0 - 1,0
good	1,01 - 2,0
moderate	2,1 - 3,0
poor	3,1 - 4,0
bad	4,01 - 5,0

Results

Distribution of periphyton

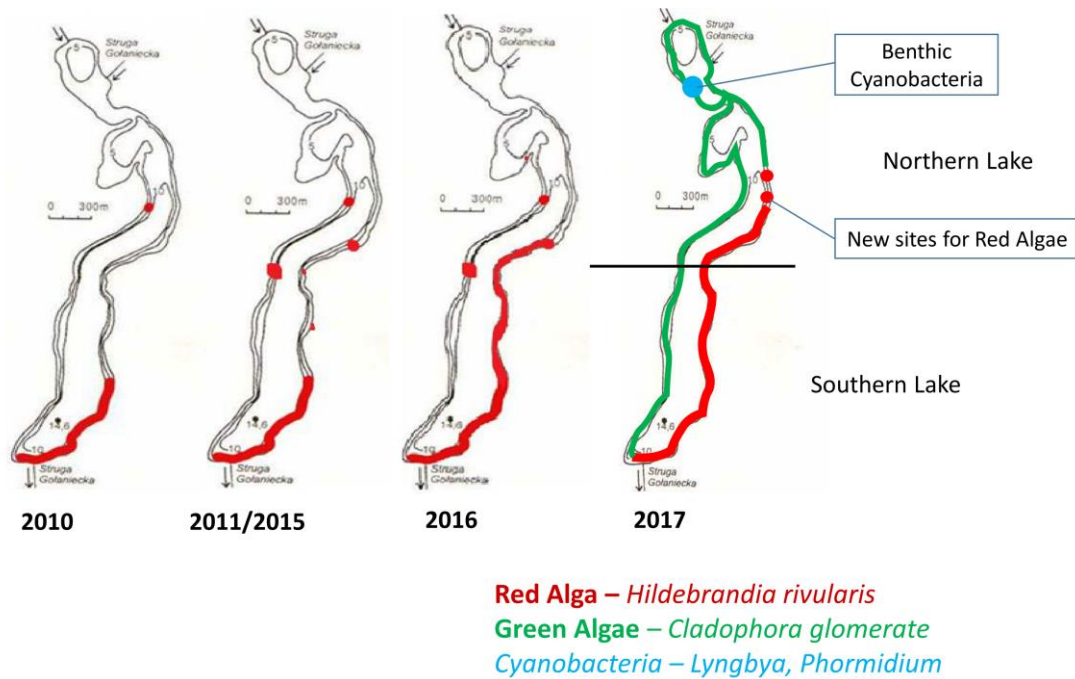


Figure 2: Distribution of periphyton in Lake Durowskie

As shown in Figure 2, the distribution of red algae (*Hildebrandia rivularis*) remains in similar spot as last year mainly in the East bank of the lake but their coverage has a northward growing pattern along the East bank. Green algae (*Cladophora glomerate*) was found in the perimeter of the lake. They have a wide range of habitat and can grow in different ecological system (Pikosz and Messyas 2016). Benthic cyanobacteria *Lyngbya* and *Phormidium* were also found in the north-west bank of the lake, sitting opposite to the inflow of the river.

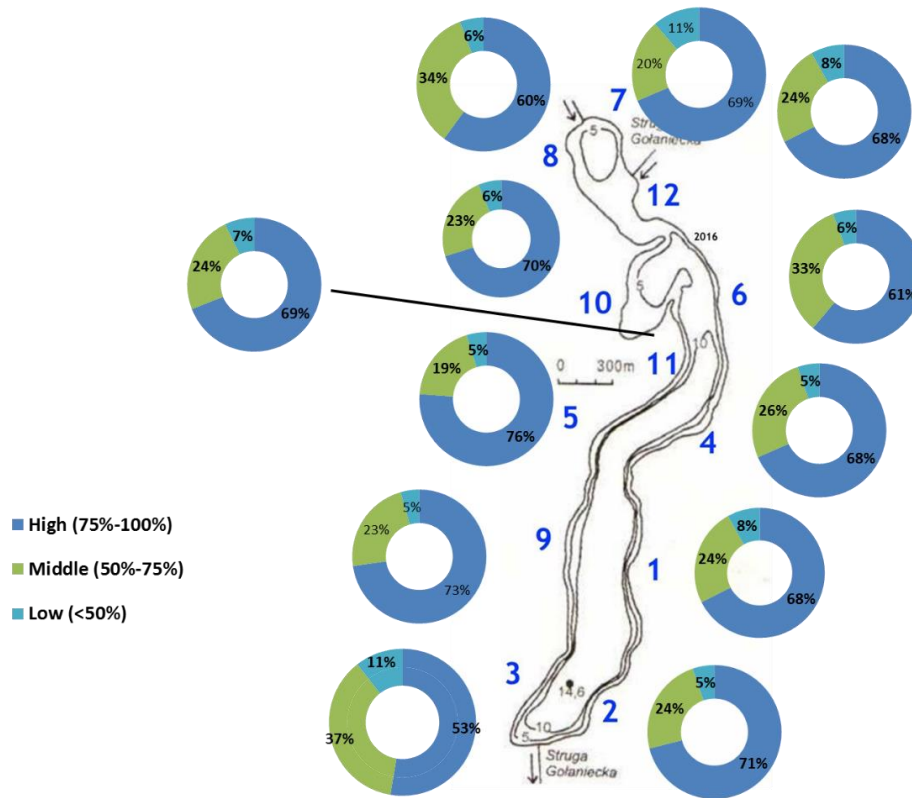


Figure 3: Periphyton Species Oxygen preference

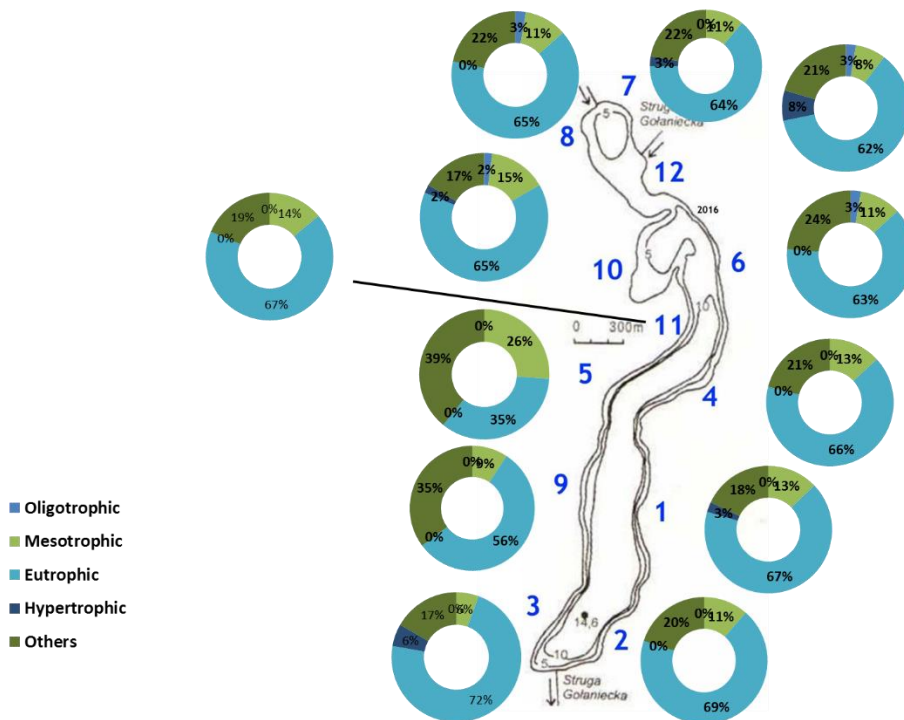


Figure 4: Periphyton Species Trophic Preference

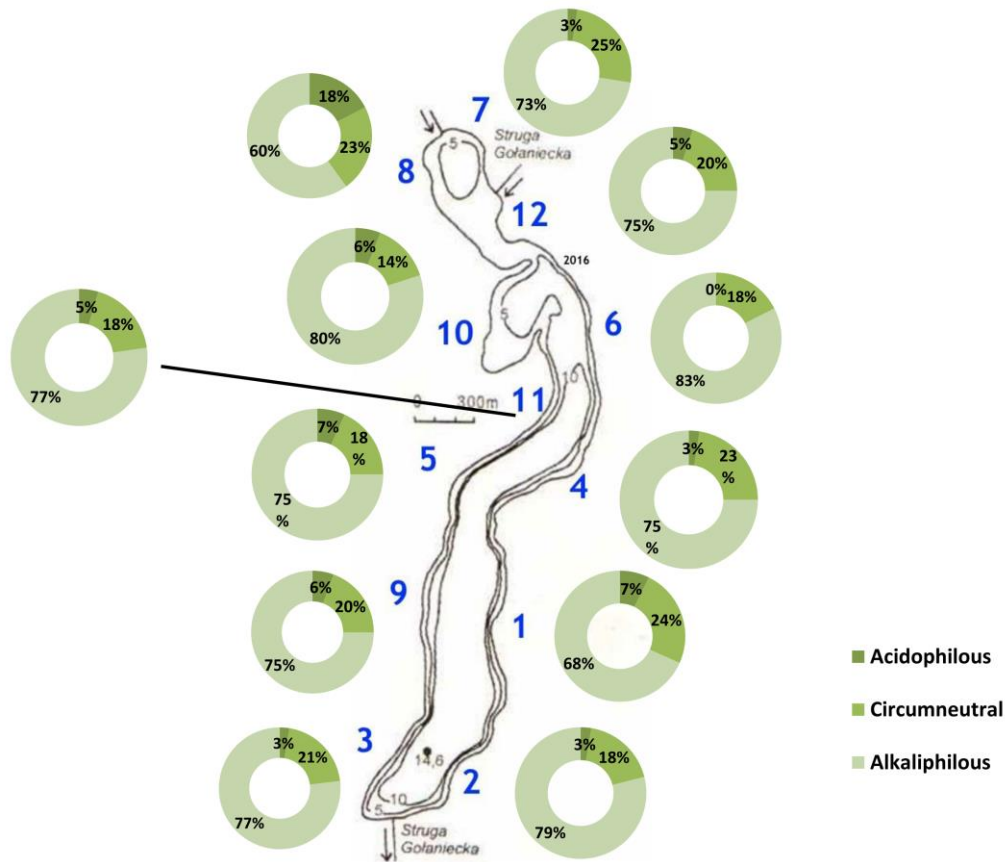


Figure 5: Periphyton Species pH Preference

Trophic Diatom Index (TDI)

The trophic diatom index shows more or less the same as it was in the last year, showing poor to moderate status of water. The remarkable drop of TDI in Site 1 (Beach 1) has shown possible negative effects from anthropogenic activities and input causing upwelling of water and mobilization of nutrients from the bottom of the water bodies. South-east part of the lake with a public beach is the most densely populated area for recreation and the TDI this year shows the area remains in a bad state of water. Still, some of the parts start getting better, implying the restoration has given positive impacts. Moreover, in Site 4, where the area has recorded increase of red algae population, shows also the greatest improvement of TDI.

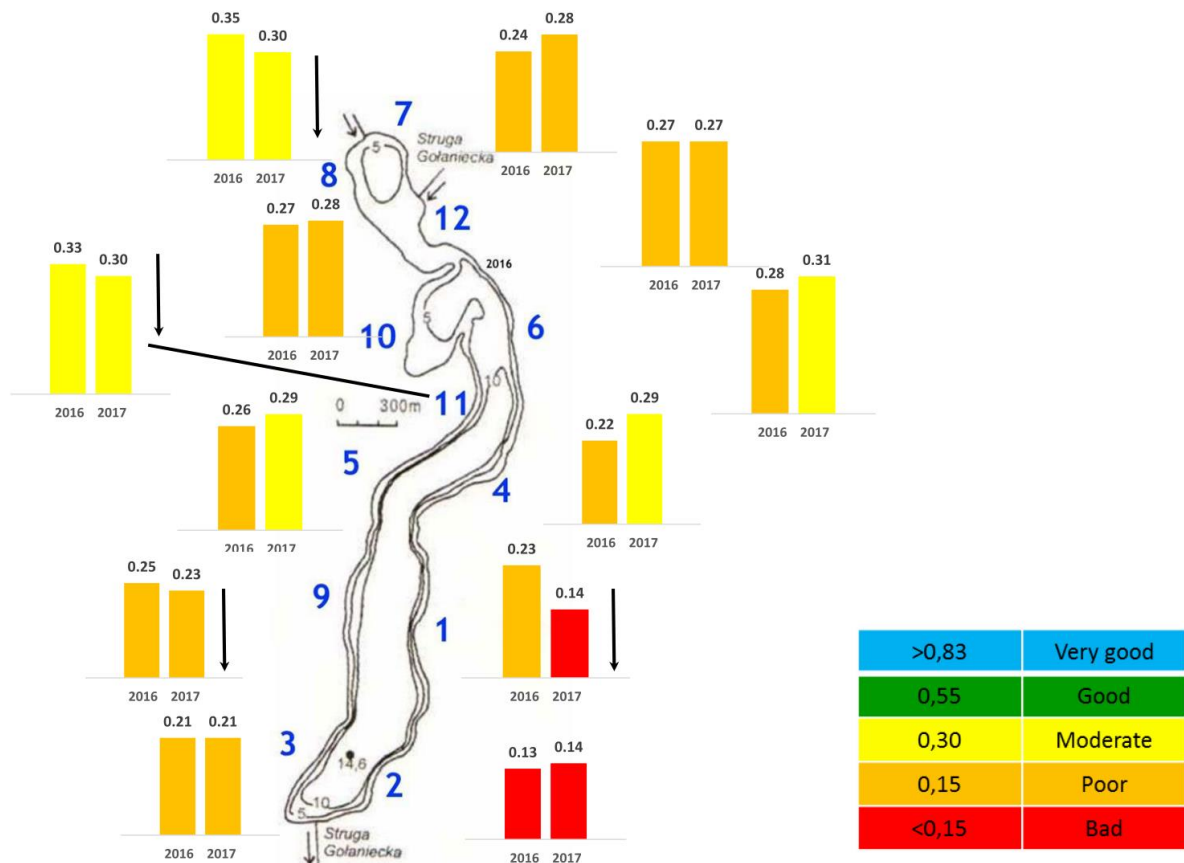


Figure 6: Trophic Diatom Index of Lake Durowskie

According to the index, the average of species is similar. Compared to the last year's result, the number of the oxygenated group increased. It means that state of littoral water improved. It occurred with almost the same number between the north and south part of the lake.

For the trophic preference, the eutrophic species still dominated in every site, compared to the last year's result, the number increased in several areas. However, from the result of pH preference, the alkaliphilous species is dominant in all parts of the lake. It could probably be a good indicator of low number of *Cyanobacteria* that can cause bloom.

Number of Species

Results – Phytoplankton

Number of species

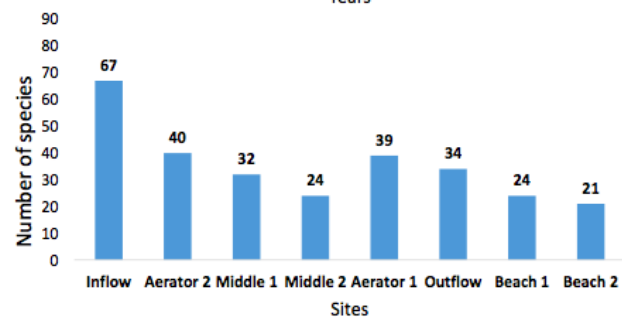
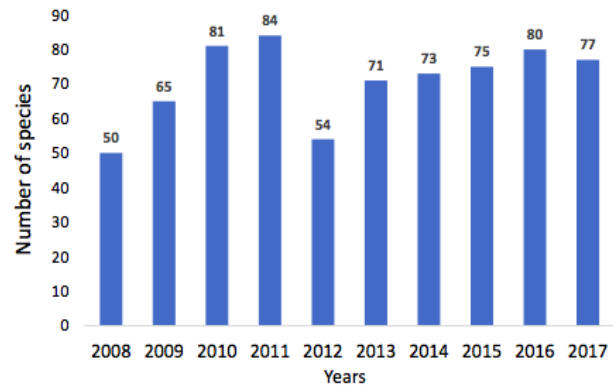
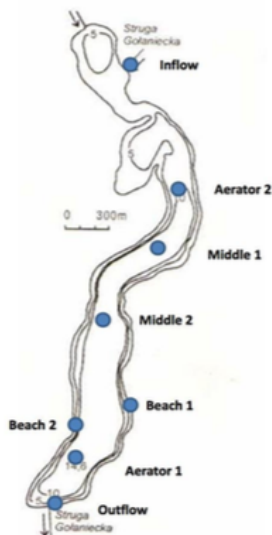


Figure 7: Number of species across years and fro individual sites

The results are interpreted in terms of year and sites respectively:

- **By year**

There are as many as 77 species found in this year. In comparison to the last year, the number of species is lower. However, if we take a look at the trends from 2013-2017, the number of species is comparable. It means that the number of species over the last five years is stabilised. We can also assume that there has been a change within the community and also shift of species, but the number remains stable.

- **Per sites**

According to the graph, the highest number of the species was concentrated at the inflow area. It is significantly different from other sites over the lake area. It can obviously be understood that the inflow is the area where there is river influence in which has carried nutrient loads from the upper stream areas. Furthermore, this area is where the turbulence occurs, so that all the sedimented part would potentially be lifted to the upper layer and later mixed with the surface layer. In addition,

weather also played important role, since during the sampling activities the weather was in rainy condition, therefore, it caused an increasing speed on current and gave implication to bigger turbulence.

Phytoplankton Diversity

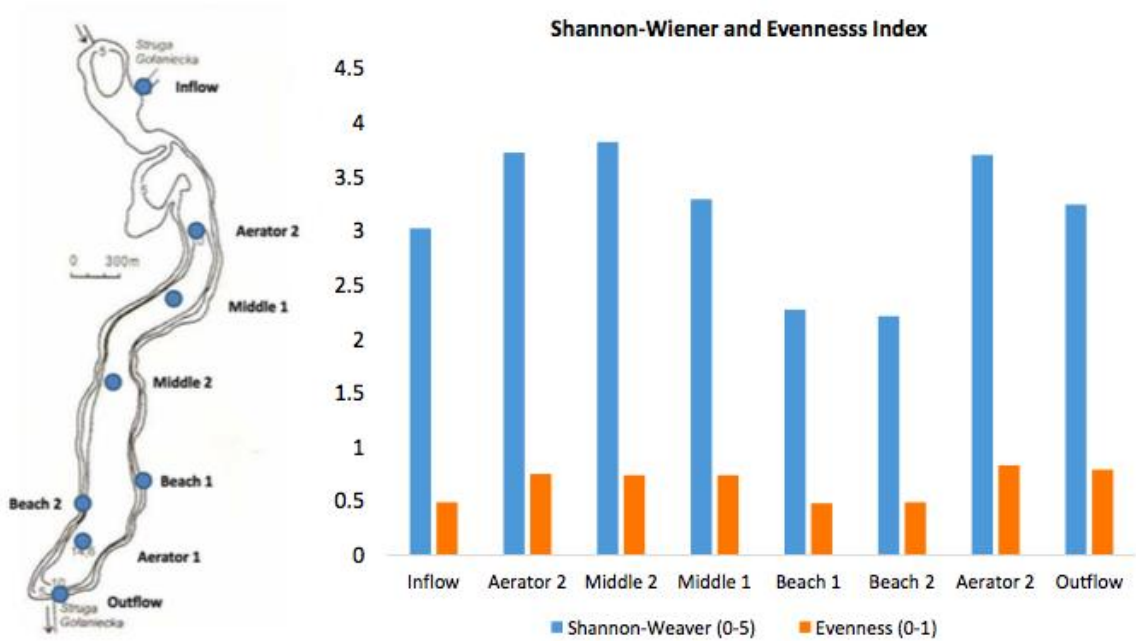


Figure 8: Species diversity and evenness of phytoplankton.

The graph shows about the distribution number of cells between taxa. According to the graph, the inflow area has very high number of species, but low number of biodiversity. It implies that there is dominant species concentrated in this area. In several parts, for example in Aerator 2, Middle 2, and Middle 1, the number of species and biodiversity is high. It shows better condition in terms of species. In contrast, in Beach 1 and Beach 2, the number of species and biodiversity is low. It could probably because those areas have high intensity of anthropogenic activities such as recreation, motor boat, and swimming activities which make some species sedimented.

Abundance of Phytoplankton groups

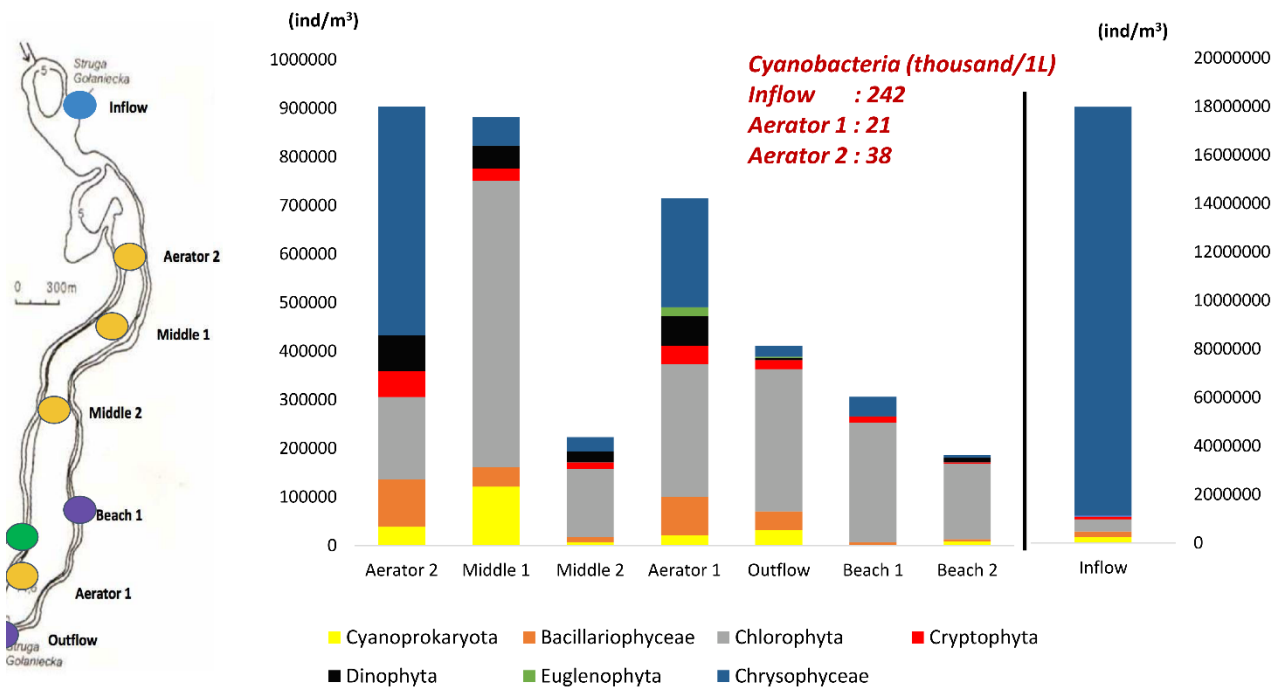


Figure 9: Abundance of phytoplankton species in Lake Durowskie.

Abundance of Phytoplankton Groups

The graphs show two different scales which compare the abundance between the groups in the lake and the groups in the river. The species which has the highest abundance was *Chrysophyta* which had almost 20 million cells/liter. It was 20 times more than those concentrated on the lake (1 million cells/liter). The reason of the abundance of this species was probably because of the late spring. It made the weather had more rainy and colder in the water area. This condition was a preference for the species of *Chrysophyta* such as *Dinobryon divergens* to thrive within such state. Furthermore, that species also has the ability to do photosynthesis, eat small bacteria, and filtrate water to increase the water visibility. There was also a high number of *Chlorophyta*. However, it was found mostly in the lake area. The other reason of this abundance of the species was also because those species built big colonies, so that the zooplankton were no able to eat them.

According to the data also, there were *Cyanobacteria* found in the river and the lake area even though only small number compared to the dominant species. There we 242/liter *Cyanobacteria* in the river. It is much higher than that of in the lake area. In addition, there were *Cyanobacteria* species like

Figure 10: Total number of individuals for phytoplankton species groups.

Anabaena flos-aqua, *Microcystis aeruginosa*, *Woronichinia nägeliana* found in the river area. They typically do not grow in Lake Durowskie area. Based on this, we can get an idea that the river is the source of Cyanobacteria, because the river carries nutrient loads from the upper stream, so that the water state in the river area is relatively highly affected by the upper stream.

Similarity Index

Table 3: Phytoplankton similarity index (Jaccard index - expressed as a percentage) over the monitoring years in Lake Durowskie.

YEAR	2009	2010	2011	2012	2013	2014	2015	2016	2017
2008	84	51	43	33	40	52	82	35	40
2009	-	48	28	20	29	35	39	13	34
2010	-	-	42	42	62	47	37	35	38
2011	-	-	-	34	58	47	50	40	38
2012	-	-	-	-	77	49	59	47	38
2013	-	-	-	-	-	52	78	45	46
2014	-	-	-	-	-	-	57	40	48
2015	-	-	-	-	-	-	-	43	47
2016	-	-	-	-	-	-	-	-	42

This index illustrates the number of species which is notably similar before and after the restoration has been done. For instance, in the first year of restoration in 2009, there was as much as 84% similar species which meant that the difference in species at that time was as much as 16%. Moving to further comparison, between 2008-2017 there is as much as 40% similar species which means that as much as 60% of species are different. It implies that after the restoration has been done for nine years, there has been an increasing number of different species. Closer look to the last year, between 2016-2017 the similarity index is 42% and the different species is 58%. It shows that there is still species shifting under the ongoing restoration.

Results – Phytoplankton

Mixed Index of Nygaard

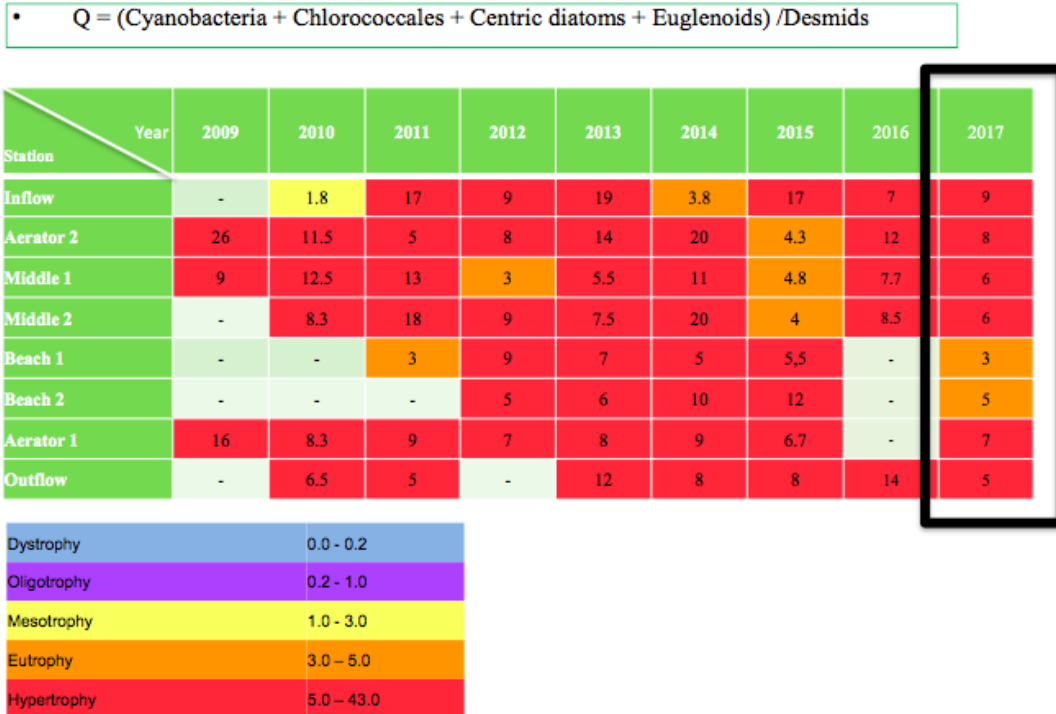


Figure 11: Mixed index from periphyton analysis

Mixed Index of Nygaard

This index was determined by the species preference to the trophic level of water. This consists of *Cyanobacteria*+*Chlorococcales* and *Centric diatoms*+*Euglenoids*/*Desmids*. According to the table, eutrophic species is still dominant, however, it is now lower compared to the last year. It means that there was an increasing number of *Desmids*. Because the higher number of *Desmids*, the lower smaller index will be. It also shows that the restoration has given improvement.

PMPL Index

Results – Phytoplankton

PMPL Index

PMPL = [YCh + YBm + YCy] / 3

where:

YCh - Chlorophyll-a concentrations

YBm - General biomass of phytoplankton

YCy - Biomass of cyanobacteria

	PMPL					
	Inflow	A2	M1	M2	A1	Outflow
2017	3.40	2.74	2.78	2.78	2.78	2.50
2016	3.70	2.78	2.76	2.76	2.67	2.53

Ecological status	PMPL
very good	0,0 - 1,0
good	1,01 - 2,0
moderate	2,1 - 3,0
poor	3,1 - 4,0
bad	4,01 - 5,0

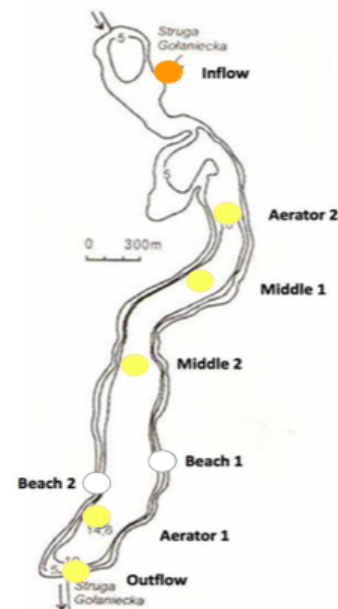


Figure 12: Trophic state of Lake Durowskie in the vertical profile indicated by PMPL index

The index is based on three different parameters (chlorophyll-a concentration, biomass of phytoplankton, and biomass of Cyanobacteria) in the vertical profile. According to the result, the data is still similar to the last year's, however, the value decreased a little bit. Overall, the water condition is in moderate state and the inflow is still in poor condition, but the value this year is lower than the last year's. In spite of the small changes, it can be concluded that the restoration has been working. Likewise, it also implies that the number of Cyanobacteria decreased. There are still Cyanobacteria found in the metalimnion, it can be taken into account for removing them in the further lake management.

Discussion

Periphyton assessment

Summarizing from periphyton measurements, the trophic state of the lake remain classified as eutrophic. Periphyton refers to organism attaching to the surface of submerged or underwater macrophytes or rocks such as algae, cyanobacteria and microbes. Compared to physical-chemical parameters, they react slower but with higher consistency to the trophic state of the lake. The results from periphyton assessment this year reflects several phenomenon. Summarizing from periphyton assessment, the species environmental preference and trophic diatom index (TDI) shows that the lake is in general under eutrophic state, alkali and oxygenated condition.

Species abundance

As shown in Figure 9 the inflow is dominated by Chrysophyceae by around 80%. When compared with results from last year, inflow area contributed to a large amount of Cyanoprokaryote. The difference might be explained by the late-spring this year when the temperature remain low in May that is not a suitable habitat for cyanobacteria, which prefers to be in warmer water. It is possible that for a delayed cyanobacteria bloom when water is warm enough for them to build colony.

Nevertheless, when looking at the amount of cyanobacteria from the inflow, Aerator 1 and Aerator 2, the count from the inflow is 242 thousands per Litre, almost 10 times the amount recorded in the aerators, implying the high amount of cyanobacterial input from the upper lake through the river. It is important to notice that even the composition of cyanobacterial in the inflow is relatively low compared to other species, its actual quantity has outweighed other areas for several times.

Bio-manipulation

By measuring the abundance of phytoplankton in different water depth, one can observe how their distribution trend develop according to their habitat preference. Water samples at 0m, 1m,

2m and 3m was been collected and the vertical distribution of Chlorophyta, Nauplius, Rotifier, Cladoera and Copepod from Aerator 1 is shown in below Figure.

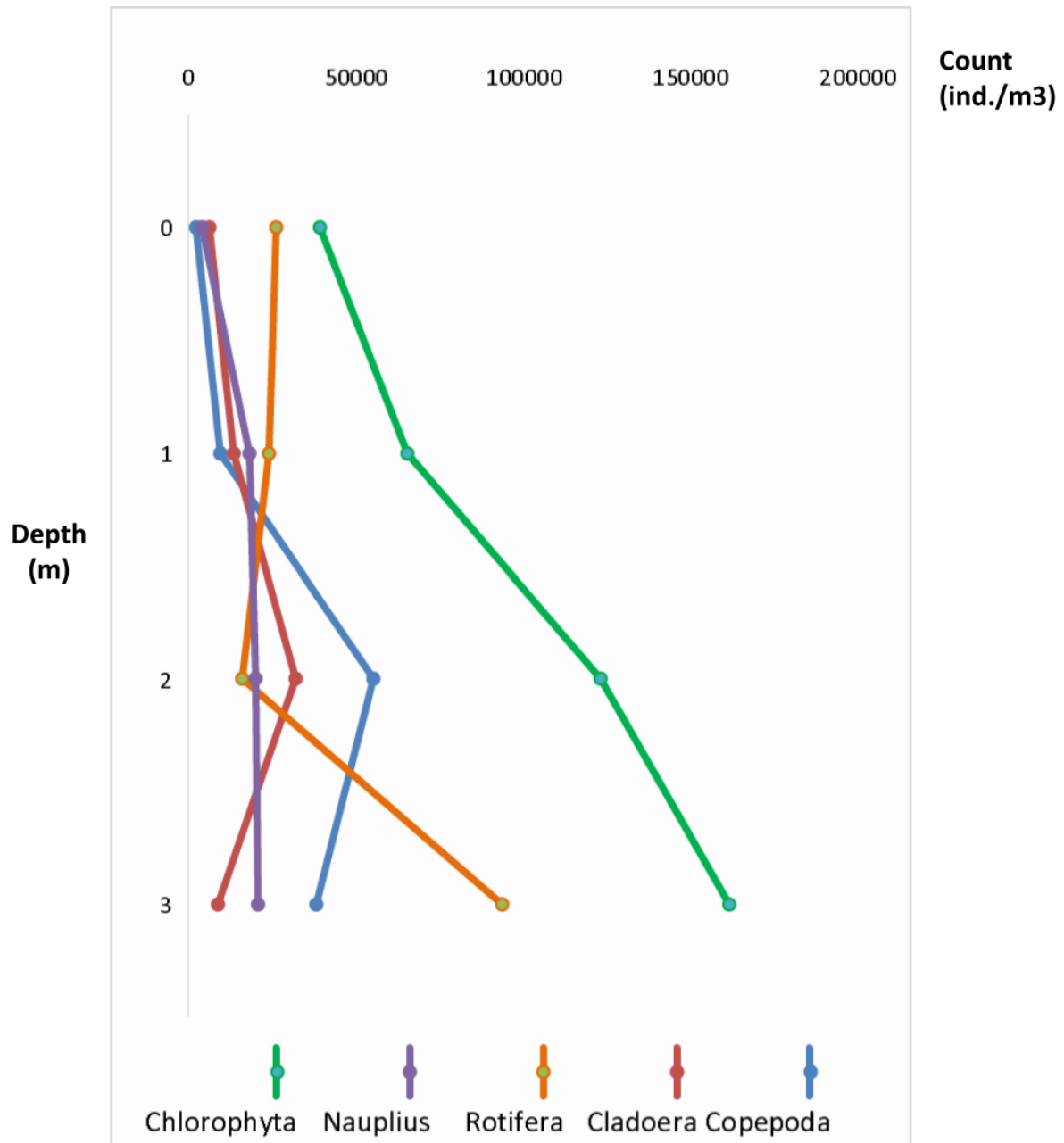


Figure 13: Vertical distribution of different phytoplankton groups at Aerator 1.

Phytoplankton and zooplankton groups occupy different water depths depending on their habitat or ecological requirements. Results of vertical distribution analysis show that there was a general decrease in the abundance of species groups at the zero meter depth. At the 1 m depth the abundance of Cladocera, Copepod and Nauplius increased slightly while that of Chlorophyta increased marginally. At the 2 m depth the abundance of all groups increased except Nauplius.

which stayed the same while that of Rotifers decreased. At the 3 m depth the abundance of Nauplius remained as previous while that of copepods and cladocera decreased. The abundance of chlorophyta increased at this depth. The general decrease in abundance of cladocera and copepods may be linked to the reduced amount of oxygen as depth increases. It was expected that as the number of zooplankton increases the number of phytoplankton should decrease due to perceived grazing pressure, however, this may not be implied in the figure above (Fig 11). We add that the species group that make up the chlorophytes are those that are able to form colonies and live in large masses. These are usually not the preferred food for zooplankton as they are difficult to consume. This is the reason why the figure shows an increasing amount of chlorophyta. This hypothesis is confirmed during identification of species as a lot of colony formed chlorophytes were found. Only a few individual cells were found signifying heavy grazing pressure on these individual cells by zooplankton. Grazing of zooplankton on phytoplankton can be confirmed.

Bio-manipulation effectiveness

By stocking the lake with pike to control perch which in a way to stabilize the amount of zooplankton was one of the restoration measures for Lake Duroske. As perch is controlled the population of zooplankton is increased to control phytoplankton. To measure the effectiveness of this restoration measure it is important to understand the vertical distribution of algae in relation to zooplankton. From the description above zooplankton density decreased at the 0 m depth but increased between the 1 and 2 meter depth. The decrease in the abundance of zooplankton at the 0 meter depth can be explained by the intensive predation by predatory fish=perch and this may be the reason for the limited abundance of zooplankton. With the availability of colony forming chlorophytes and higher abundance of this groups supported by data we confirm the effective grazing on single cell chlorophyta. This confirms the improvement of Lake condition through bio-manipulation.

Highlights from 2017

Enhancement in Species diversity and new species

Despite of the slight decrease number of species discovered this year, there are four new species found in the lake. As shown by Shannon-Wiener Index and Evenness index, the species

diversity improved for most of the site. Most of the species were found in the inflow of the lake because the inflow consists of nutrient-rich water, for example, number of diatom species found in the inflow and water bodies of the lake is 28 and 8 respectively.

Shifting of cyanobacteria

Cyanobacteria has been the major concern of the lake and benthic cyanobacteria was discovered this year at the north-west bank of the lake at a position opposite to the inflow point, as shown in Figure 6. It is possible that under the effect of wind and current, the influx of cyanobacteria from inflow being flushed towards the north-west bank. Besides, the distribution of cyanobacteria shows downward movement trend to aerators. It might be due to the upwelling of water by waves induced by wind and anthropogenic recreational activities.

Northward spreading of red algae

Red algae *H. rivularis*, mainly inhabited eutrophic and alkali water (Eloranta and Kwandrans 2004), has been classified as an indicator for good water quality state (Gutowski et al. 2004). Red algae existed in Lake Durowskie for several years and their coverage kept growing every year. They have a northward spreading trend along the lake bank, indicating good water quality and eutrophic state in the area. The distribution of red algae can be explained by their preference to shaded area (sensitive to sunlight), alkali and oxygenated water and under the wind direction of the lake from the West, inducing current to push red algae to the western bank of the lake.

Similar species preference in North and South

By using species composition, the oxygen preference, trophic preference and pH preference of the species show the lake water state as a bio-indicator. After comparing results from the northern and southern part of the lake, they are found to have very similar results, as shown in (Figure 12). The result implies that the water from the north and the south tends to be oxygenated, alkali and eutrophic.

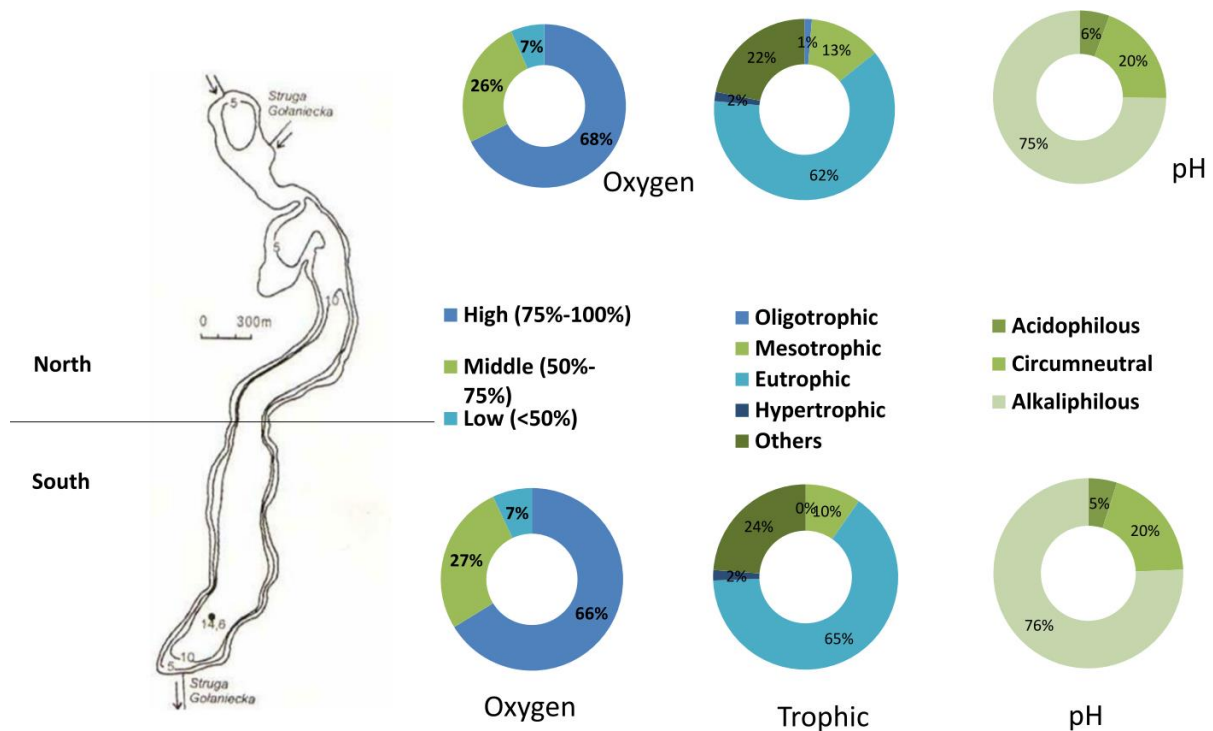


Figure 14: Summary of species preference between north and south part of the Lake Durowskie

Reduced effectiveness in bio-manipulation

The practice of restocking pike has implemented for several years in order to recover the top-down control mechanism in the lake. Although last year recorded a positive result on bio-manipulation, this year has become less effective due to the delay of the restocking and the size of the restocking pike is too small to sustain in the control mechanism.

Conclusion

To sum up, different parameters from algal species supported that the lake is still in eutrophic status but showing improvements from last year. Existence of red and green algae as well as the environmental preference of periphyton indicate improving water. Nevertheless, the influx from upper lake Struga Gołaniecka, which contains lots of nutrients such as nitrogen and phosphorus remains a problem as this year cyanobacteria was found at the northwest of the lake, a site parallel to the inflow. Moreover, the effect of human activities concentrated in the southern part of the lake also indicated by a noticeable drop of diatom index in the Beach 1 position. On the other hand, with four new species found in this year, the diversity of phytoplankton species also improved significantly from last year. The PMPL indicates a

moderate trophic state for Lake Durowskie and there are also evidence for effective grazing of zooplankton towards phytoplankton where relatively greater portions of zooplankton contributes to Rotifers. Due to the growing colony formed by chlorophyta which is too big to be preyed by zooplankton, a large amount of chlorophyta are found in deeper water and hence, further control for phytoplankton is suggested to maintain effective top-down control mechanism in the lake. Finally, bio-manipulation has shown lower effectiveness due to the delay schedule of the pike stocking and the inappropriate size of the pike. As a result, coordination and mutual efforts between different working parties are critical to the status of the lake and remedial measures shall keep implementing to ensure the stable water quality for sustaining tourism and human activities in the lake.

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Annexes

ANNEX 1. Comparison of phytoplankton species composition in different investigated years in June/July in Lake Durowskie.

Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
Cyanoprokaryota – cyanobacteria										
<i>Anabaena affinis</i> Lemm.							+		+	
<i>Anabaena flos-aquae</i> Brebisson		+				+			+	+
<i>Aphanizomenon aphanizomenoides</i> (Forti) Hort. & Kom.	+				+					
<i>Aphanizomenon flos-aquae</i> (L.) Ralfs	+	+	+	+			+	+		+
<i>Aphanizomenon gracile</i> Lemmerman	+									
<i>Aphanizomenon isatschenkoi</i> (Usacc.) Pros. - Lavrenko	+	+	+							
<i>Aphanocapsa grevillei</i> (Ber.) Rabenhorst		+								
<i>Aphanocapsa incerta</i> (Lemm.) Cronberg et Komarek	+	+	+			+				+
<i>Arthrospira massartii</i> Kuff.		+								
<i>Chroococcus limneticus</i> Lemm.	+	+		+						+
<i>Chroococcus turgidus</i> (Kütz.) Naeg.		+		+			+	+		+
<i>Cyanogranis feruginea</i> (Wawrik) Hind.		+	+							
<i>Gloeocapsa minuta</i> Lemm.							+			+
<i>Jaaginema pseudogeminatum</i> (Schmid) Anagn. et Kom.			+	+						

Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Limnothrix lauterbornii</i> (Schmidle) Anagn.		+								
<i>Limnothrix redekei</i> (Van Goor) Meffert	+		+	+	+	+	+	+	+	+
<i>Lyngbya hieronymusii</i> Lemm.		+						+		
<i>Microcystis aeruginosa</i> Kützing	+			+	+	+	+	+	+	+
<i>Microcystis flos-aquae</i> (Wittrock) Kirchner			+			+				
<i>Jaaginema gracils</i> (Bocher) Anagn. et kom.		+						+		
<i>Phormidium granulatum</i> Gardn. Anagn.	+	+	+		+					
<i>Phormidium tenue</i> (Agards ex Gomont) Anagn. et kom.		+								+
<i>Phormidium autumnale</i> Gomont									+	+
<i>Planktolyngbya limnetica</i> (Lemm.) Kom. – Legn. Et Cronenberg		+	+	+	+	+	+	+	+	+
<i>Planktothrix agardhii</i> (D.C. ex Gom.) Anagn. Et Kom.	+	+	+	+	+	+	+	+	+	+
<i>Pseudanabaena limnetica</i> (Lemm.) Kom.	+	+	+				+	+		+
<i>Woronichina naegeliana</i> (Unger)Elenkin								+	+	+
<i>Spirulina laxissima</i> (W. West)			+							
<i>Spirulina maior</i> Kütz.					+					
<i>Oscillatoria grossegranulata</i> Skuja						+				
Bacillariophyceae – diatoms										
<i>Achnanthes exigua</i> Grun.			+							

<i>Achnanthes minutissima</i> Kützing	+		+	+		+		+		+
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Amphora copulate</i> (Kütz.) Schoeman & Archibald								+		
<i>Amphora ovalis</i> Kützing	+	+	+	+						+
<i>Amphora pediculus</i> (Kütz.) Grun.			+							
<i>Asterionella formosa</i> Hasall		+		+	+	+	+	+	+	+
<i>Caloneis amphisbaena</i> (Bory) Cleve								+		
<i>Cocconeis euglypta</i> (Ehr.) Clevei	+									
<i>Cocconeis pediculus</i> Ehr.										+
<i>Cocconeis placentula</i> Ehr.	+		+	+	+	+		+	+	+
<i>Cyclotella atomus</i> Hustedt		+								
<i>Cyclotella meneghiniana</i> Kütz.	+	+	+			+				+
<i>Cyclotella ocellata</i> Pant.	+		+	+	+	+	+	+	+	+
<i>Cyclotella operculata</i> (Ag.) Kützing	+	+	+			+				+
<i>Cyclotella radiosa</i> (Grun.) Lemm.	+	+	+	+	+	+	+	+	+	+
<i>Cymatopleura solea</i> (Breb.) W. Smith							+			+
<i>Cymbella affinis</i> Kützing				+						
<i>Cymbella amphicephala</i> Naegeli ex Kütz.										+
<i>Cymbellalanceolata</i> (Ehr.) Kirchner										+

<i>Cymbella microcephala</i> Grun.				+		+				
<i>Cymbella minuta</i> Hilse ex Rabenhorst	+		+	+	+	+	+	+	+	+
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Diatoma vulgare</i> Bory				+				+		
<i>Fragilaria capucina</i> (Desm.) Rabenhorst				+		+				
<i>Fragilaria crotonensis</i> Kitton	+	+		+	+	+	+	+	+	+
<i>Fragilaria pinnata</i> Ehr.	+			+			+	+		
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot	+	+	+	+	+	+	+	+	+	+
<i>Fragilaria ulna</i> var. <i>Angustissima</i> Sippen	+	+	+	+	+	+	+	+		+
<i>Gomphonema acuminatum</i> Ehr.					+	+	+			
<i>Gomphonema olivaceum</i> (Horn.) Breb.			+	+			+			+
<i>Gomphonema parvulum</i> (Kütz.) Kütz.				+					+	
<i>Melosira varians</i> Ag.	+									
<i>Hippodonta capitata</i> (Ehr.) L-B, Metz. et Witk.						+		+	+	
<i>Navicula cincta</i> (Ehr.) Ralfs	+	+	+			+	+	+		+
<i>Navicula mensiculus</i> Schumann	+									
<i>Navicula radiosa</i> Kützing			+	+	+	+	+		+	+
<i>Navicula lanceolata</i> Ehr.									+	
<i>Naviula tripunctata</i> (O.F. Muller) Bory de Sain. Van.			+			+		+		+

<i>Nitzschia palea</i> (Kütz.) W. Smith				+	+	+	+	+		
<i>Nitzschia recta</i> Hantzsch ex Rabenh.						+				
<i>Nitzschia sigmaidea</i> (Ehr.) W. Smith				+			+	+		
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Nitzschia sinuata</i> (W. Sm.) Grunow				+						
<i>Pinnularia maior</i> (Kütz.) Rabenhorst									+	
<i>Pinnularia viridis</i> (Nitzsch) Ehr.				+			+	+		
<i>Placoneis gastrum</i> (Ehr.) Meresch.		+								
<i>Rhopalodia gibba</i> (Ehr.) Muller						+		+		+
<i>Staurosira construens</i> Ehr.		+							+	
Chlorophyta - green algae										
<i>Actinastrum hantzschii</i> Lagerh.									+	
<i>Ankistrodesmus falcatus</i> (Corda) Ralfs		+								+
<i>Botryococcus braunii</i> Kütz.						+	+	+	+	+
<i>Characium aqngustatum</i> A. Braun		+		+	+	+	+	+	+	
<i>Chlamydomonas globosa</i> Snow	+	+	+	+		+				
<i>Chlamydomonas passive</i> Skuja			+			+	+	+		
<i>Chlamydomonas reinhardtii</i> Dangeard		+						+		
<i>Closteriumacutum</i> var. <i>variabile</i> (Lemm.) Krieg.	+		+	+				+	+	+

<i>Coelasrum astroideum</i> De Notaris			+	+	+	+		+	+	+
<i>Coelastrum microporum</i> Naegel.			+							+
<i>Coelastrum reticulatum</i> (Dang.) Senn	+	+					+		+	+
<i>Coenocystis planctonica</i> Korshikov									+	
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Cosmarium abbreviatum</i> Raciborski	+		+	+	+	+	+			
<i>Cosmarium exiguum</i> W. Archer		+								
<i>Cosmarium formulosum</i> Lund								+		
<i>Cosmarium trilobulatum</i> Reinsh								+		
<i>Cosmarium margaritatum</i> (Turp.) Ralfs				+					+	
<i>Cosmarium phaseolus</i> Brebisson in Ralfs	+		+	+		+	+	+	+	
<i>Cosmarium leave</i> Rabenhorst					+					
<i>Cosmarium regnellii</i> Wille	+	+	+		+		+	+	+	+
<i>Crucigeniella rectnagulrais</i> (Naeg.) Kom.						+				
<i>Crucigenia tetrapedia</i> (Kirchner) W. et G.S. West			+							+
<i>Desmodesmus communis</i> (Hegew.) Hegew.	+	+	+	+	+	+	+	+	+	+
<i>Desmodesmus grahneisii</i> (Heynig) Fott				+						
<i>Desmodesmus naegellii</i> (Meyen) Hegew.			+							
<i>Desmodesmus opoliensis</i> (Richter) Hegew.			+			+				

<i>Desmodesmus subspicatus</i> (Chod.) Hegew. et Schmidt	+		+			+	+	+		
<i>Dicellula geminate</i> (Printz) Kors.									+	
<i>Dictyosphaerium pulchellum</i> Wood	+	+	+	+						+
<i>Didymocystis planctonica</i> Korsikov				+		+				
<i>Elkatothrix gelatinosa</i> Wille			+	+		+	+	+	+	+
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Franceia ovalis</i> (France) Lemm.			+				+			
<i>Golenkinia radiata</i> Chodat	+		+	+	+	+	+	+	+	
<i>Kirchneriella contorta</i> var. <i>elegans</i> (Schmidle) Bohlin	+					+	+	+		+
<i>Kirchneriella incurvata</i> Belcher et Swale									+	
<i>Kirchneriella obesa</i> (West) West & West									+	
<i>Koliella longiseta</i> (Vischer) Hindak	+									
<i>Lagerheimia ciliata</i> (Lag.) Chodat							+			
<i>Micractinium crassisetum</i> Hortobagyi				+						
<i>Micractinium pusillum</i> Fresenius				+					+	
<i>Mougeotia</i> sp.				+	+					
<i>Monoraphidium arcuatum</i> (Kors.) Hindak	+									
<i>Monoraphidium circinale</i> (Nyg.) Nygaard	+									
<i>Monoraphidium contortum</i> (Thur.) Kom.-Legn.	+	+	+	+	+	+	+	+	+	+

<i>Monoraphidium griffithii</i> (Berk.) Kom.-Legn.	+		+				+	+		
<i>Monoraphidium irregulare</i> (G.M. Sm.) Kom.-Legn.	+		+							
<i>Monoraphidium komarkovae</i> Nygaard	+	+	+				+	+		+
<i>Monoraphidium minutum</i> (Nageli) Kom. - Legn.		+								
<i>Monoraphidium obtusum</i> (Kors.)Kom. - Legn.	+									
<i>Nephrocytiuma gardhianum</i> Naegeli									+	
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Nephrocytium limneticum</i> (G. M. Sm.) G. M. Sm.				+						
<i>Oocystis lacustris</i> Chodat	+	+	+	+	+		+	+	+	+
<i>Oedogonium sp.</i>								+		+
<i>Palmelochette tenerrima</i> Kors.				+						
<i>Pandorina morum</i> (O.F. Müller) Bory			+			+				+
<i>Pediastrum biradiatum</i> Meyen									+	
<i>Pediastrum boryanum</i> (Turpin) Meneg.			+	+	+	+	+	+	+	+
<i>Pediastrum simplex</i> Meyen						+			+	+
<i>Pediastrum duplex</i> Meyen						+	+	+	+	+
<i>Pediastrum duplex var. gracillium</i> West									+	
<i>Pediastrum tetras</i> (Ehr.) Ralfs			+				+			
<i>Phacotus lendneri</i> Chodat.				+	+					

<i>Phacotus lenticularis</i> (Ehr.) Stein	+			+	+	+	+	+	+	+
<i>Plankosphaceeria gelatinosa</i> Meyen									+	
<i>Provasoliella saccata</i> (Skuja) Ettl					+					
<i>Provasiorella</i> sp.							+			
<i>Pteromonas angulosa</i> (Carter) Lemm.		+	+						+	
<i>Pteromonas cordiformis</i> Lemm.			+						+	
<i>Scenedesmus cuminatus</i> (Lager.) Chodat			+		+		+	+	+	+
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Scenedesmus bicaudatus</i> Dedusenko			+	+	+				+	
<i>Scenedesmus dimorphus</i> (Turp.) Kütz.		+		+						
<i>Scenedesmus ecornis</i> (Ehr.) Chod.			+	+	+		+		+	
<i>Scenedesmus obtusus</i> Meyen				+						+
<i>Scenedesmus regularis</i> Swirenko		+								
<i>Scenedesmus verucosus</i> Roll				+						
<i>Sphaerocystis planctonica</i> (Korsikov) Bourrelly				+	+	+	+	+	+	+
<i>Staurastrum gracile</i> Ralfs			+	+	+	+	+	+	+	+
<i>Staurastrum paradoxum</i> Meyen							+			
<i>Staurastrum tetracerum</i> Ralfs ex Ralfs									+	+
<i>Tetraedron caudatum</i> (Corda) Hansgirg	+		+					+		

<i>Tetraedron minimum</i> (A. Br.) Hansgirg	+	+	+	+	+	+	+	+	+	+
<i>Tetraedron triangulare</i> (Chod.) Kom.	+	+		+		+			+	
<i>Tetrastrum glabrum</i> (Roll) Ahlstr. et Tiff			+	+			+			+
<i>Tetrastrum staurogeanieforme</i> (Schroed.) Lemm.			+	+		+	+	+	+	+
<i>Treubaria schmidlei</i> (Schroeder) Fott et Kovacik		+	+	+		+	+			
<i>Cryptophyta - cryptophytes</i>										
<i>Chroomonas acuta</i> Uterm.	+				+		+			
<i>Cryptomonas erosa</i> Ehrenberg	+	+	+	+	+	+	+	+	+	+
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Cryptomonas gracilis</i> Skuja		+								
<i>Cryptomonas marssonii</i> Skuja	+	+	+	+	+	+	+	+	+	+
<i>Cryptomonas ovata</i> Ehrenberg	+	+	+	+	+	+	+	+	+	+
<i>Cryptomonas rostrata</i> Troitzskaja emend I. Kiselev	+		+	+	+	+	+	+	+	+
<i>Rhodomonas minuta</i> Skuja	+	+	+	+	+	+	+	+	+	+
<i>Dinophyta - dinophytes</i>										
<i>Ceratium hirundinella</i> (F. B. Müller) Bergh	+	+		+	+	+	+	+	+	+
<i>Ceratium cornutum</i> (Ehr) Clap.& Lachman								+		
<i>Gymnodinium aeruginosum</i> Stein	+					+		+		
<i>Peridiniopsis cuningtonii</i> Lemm.	+	+	+	+	+	+	+	+	+	+

<i>Peridinium cinctum</i> (O.F. Müller) Ehrenberg	+	+	+	+	+	+	+	+	+	+
<i>Peridinium gatunense</i> Nygaard						+				
<i>Peridinopsis berolinense</i> (Lemm.) Bourrelly	+	+	+	+	+	+	+	+	+	+
<i>Peridinopsis elpatiewskyi</i> (Ostenf.) Bourrelly	+		+	+	+	+	+	+	+	+
<i>Peridinopsis kevei</i> Grig. & Vasas									+	
<i>Euglenophyta - euglenoids</i>										
<i>Colacium vesiculosum</i> Ehr.		+		+		+		+		
<i>Euglena caudata</i> Hübner					+				+	
<i>Euglena pisciformis</i> Klebs		+		+				+	+	
Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017
<i>Phacus caudatus</i> Hubner								+		
<i>Phacus pusillus</i> Lemm.								+	+	
<i>Phacus orbicularis</i> Hubner	+	+								
<i>Trachelomonas hispida</i> (Perty) Stein	+		+	+	+	+	+	+	+	+
<i>Trachelomonas intermedia</i> Dangeard									+	
<i>Trachelomona splanctonica</i> Swirenko	+		+	+						
<i>Trachelomonas volocina</i> Ehrenberg	+		+	+	+	+	+		+	
<i>Chrysophyceae - chrysophytes</i>										
<i>Chrysococcus rufescens</i> Klebs			+	+						

<i>Dinobryon bavaricum</i> Imhoff		+	+	+	+	+	+		+	
<i>Dinobryon crenulatum</i> W. et G.S. West		+	+	+						
<i>Dinobryon divergens</i> Imhof		+	+		+	+	+	+	+	+
<i>Dinobryon sociale</i> Ehrenberg	+		+	+		+			+	
<i>Erkenia subaequiciliata</i> Skuja	+	+	+		+	+	+	+	+	+

ANNEX 2. List of phytoplankton species from different taxonomical algal groups and their frequency in Lake Durowskie from 26th June to 30th June 2017 (Inf – inflow; A2 – Aerator 2; Mid. 2 – Middle 2; Mid.1 – Middle 1; Outf – outflow; B1 – Beach 1; B2 – Beach 2; F – frequency).

	n=1	n=4	n=4	n=4	n=4	n=2	n=1	n=1	n=21
Depth	0m	0-3m	0-3m	0-3m	0-3m	0-1m	0m	0m	%
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2	F
Cyanoprokaryota - cyanobacteria									
<i>Aphanocapsa incerta</i> (Lemm.) Cronberg et Komarek	+	+	+	+	+	+		+	52
<i>Anabaena lemmermanii</i> P. Richter	+								5
<i>Anabaena flos-aquae</i> Brebisson	+								5

<i>Aphanizomenon flos-aquae</i> (L.) Ralfs	+	+	+						29
<i>Chroococcus turgidus</i> (Kütz.) Naeg.							+		5
<i>Limnithrix redekei</i> (Van Goor) Meffert	+	+	+	+	+	+		+	71
<i>Microcystis aeruginosa</i> Kützing	+	+							10
<i>Merismopedia punctata</i> Meyen							+		5
<i>Chroococcus limneticus</i> Lemm.		+							5
<i>Planktolyngbya limnetica</i> (Lemm.) Kom. – Legn. Et Cronenberg		+				+		+	14
<i>Planktothrix agardhii</i> (D.C. ex Gom.) Anagn. et Kom.	+	+	+	+	+				48
<i>Pseudanabaena limnetica</i> (Lemm.) Kom.		+				+		+	14
<i>Woronichina naegeliana</i> (Unger) Elenkin	+								5
Bacillariophyceae- diatoms									
<i>Amphora ovalis</i> Kützing	+					+		+	14
<i>Achnanthes minutissima</i> Kützing					+				10
<i>Asterionella formosa</i> Hasall	+								5
<i>Cocconeis placentula</i> Ehr.	+								5
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2	F
<i>Cocconeis pediculus</i> Ehr.	+								5
<i>Cyclotella ocellata</i> Pant.	+	+	+	+	+	+			76
<i>Cyclotella radiosa</i> (Grun.) Lemm.	+	+	+	+	+	+	+		71
<i>Cyclotella meneghiniana</i> Kützing	+								5
<i>Cymbella amphicephala</i> Grun.	+								5
<i>Cymbella lanceolata</i> (Ehr.) Kirchner	+								5
<i>Cymbella minuta</i> Hilse ex Rabenhorst	+	+		+	+	+	+		38
<i>Cymatopleura solea</i> (Breb.) W. Smith	+								5
<i>Epithemia sorex</i> Kützing	+								5

<i>Fragilaria construens</i> (Ehr.) Grunow	+								5
<i>Fragilaria crotonensis</i> Kitton	+	+			+	+	+		29
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot	+	+	+	+	+	+		+	52
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen	+					+			10
<i>Gomphonema olivaceum</i> (Horn.) Breb.	+								5
<i>Hippodonta capitata</i> (Ehr.) L-Bert., Metz. & Witkowski						+			5
<i>Meridion circulare</i> (Grev.) C. Agardh	+		+						10
<i>Navicula cincta</i> (Ehr.) Ralfs	+					+			10
<i>Navicula lanceolata</i> (Agardh) Ehr.	+								5
<i>Navicula radiosa</i> Kützing	+	+			+	+	+	+	38
<i>Navicula reinhardtii</i> Grun.	+								5
<i>Naviula tripunctata</i> (Muller) Bory	+					+			10
<i>Nitzschia palea</i> (Kütz.) W. Smith	+								5
<i>Nitzschia recta</i> Hantzsch	+								5
<i>Nitzschia sigmoidea</i> (Ehr.) W. Smith	+								5
<i>Pinnularia viridis</i> (Nitzsch) Ehr.	+								5
<i>Rhopalodia gibba</i> (Ehr.) Muller	+								5
<i>Stephanodiscus astrea</i> (Ehr.) Grunow		+							5
Chlorophyta- green algae									
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2	F
<i>Botryococcus braunii</i> Kutzing	+	+			+	+			29
<i>Cladophora glomerata</i> (L.) Kutzing			+						5
<i>Closterium acutum</i> var. <i>variabile</i> (Lemm.) Krieg.		+							5
<i>Coelastrum astroideum</i> De Notaris	+	+	+	+	+	+	+	+	71
<i>Coelastrum microporum</i> Naegel.	+	+		+					19
<i>Coelastrum reticulatum</i> (Dang.) Senn	+	+	+		+	+			33

<i>Cosmarium regnellii</i> Wille	+		+						10
<i>Crucigenia tetrapedia</i> (Kirchner) W. et G.S. West			+						10
<i>Desmodesmus communis</i> (Hegew.) Hegew.	+	+			+		+		19
<i>Dictyosphaerium pulchellum</i> Wood	+								5
<i>Elkatothrix gelatinosa</i> Wille	+				+	+	+		29
<i>Eudorina elegans</i> Ehr.				+			+		10
<i>Eutetramorus planctonicus</i> (Kors.) Bourr.	+	+	+	+	+	+	+	+	95
<i>Kirchneriella contorta</i> var. <i>elegans</i> (Schm.) Bohlin	+	+			+				14
<i>Monoraphidium contortum</i> (Thur.) Kom.-Legn.	+	+	+		+				19
<i>Oocystis lacustris</i> Chodat	+	+	+	+	+	+		+	76
<i>Pediastrum boryanum</i> (Turpin) Meneg.	+	+	+	+	+		+	+	71
<i>Pediastrum duplex</i> Meyen	+	+	+	+	+	+	+		52
<i>Phacotus lenticularis</i> (Ehr.) Stein	+	+	+	+	+	+	+	+	90
<i>Scenedesmus acuminatus</i> (Lager.) Chodat	+	+			+				14
<i>Scenedesmus obtusus</i> Meyen								+	5
<i>Sphaerocystis planctonica</i> (Korsikov) Bourrelly	+	+	+	+	+	+	+	+	95
<i>Tetraedron minimum</i> (A. Br.) Hansgirg	+	+			+				19
<i>Tetrstrum komarekii</i> Hindak			+						5
<i>Tetrastrum staurogeanieforme</i> (Schroed.) Lemm.	+		+		+	+	+		29
<i>Staurastrum gracile</i> Ralfs	+	+	+		+	+	+		57
Cryptophyta- cryptophytes									
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2	F
<i>Cryptomonas erosa</i> Ehrenberg	+	+	+	+	+	+	+	+	76
<i>Cryptomonas marssonii</i> Skuja	+		+		+			+	19
<i>Cryptomonas ovata</i> Ehrenberg							+		5
<i>Cryptomonas rostrata</i> Troitz. emend I. Kiselev	+	+	+	+	+	+	+	+	67
<i>Rhodomonas minuta</i> Skuja	+	+	+	+	+	+	+		67

Dinophyta- dinophytes									
<i>Peridiniopsis berolinense</i> (Lemm.) Bourrelly	+		+	+	+			+	33
<i>Ceratium hirundinella</i> (F. B. Müller) Bergh	+	+	+	+	+	+	+		67
<i>Peridiniopsis cuningtonii</i> Lemm.	+	+	+	+	+	+	+	+	76
<i>Peridinium cinctum</i> (O.F. Müller) Ehrenberg	+	+	+	+	+	+		+	81
<i>Peridiniopsis elpatiewskyi</i> (Ostenf.) Bourrelly	+	+		+	+				33
Euglenophyta- euglenoids									
<i>Trachelomonas hispida</i> (Perty) Stein	+		+		+				24
<i>Euglena acus</i> Ehr.					+				10
<i>Colacium vesiculosum</i> Ehr.						+			5
Chrysophyceae- chrysophytes									
<i>Erkenia subaequiciliata</i> Skuja	+	+	+	+	+	+	+	+	86
<i>Dinobryon divergens</i> Imhof	+	+	+		+	+	+	+	76

ANNEX 3. Average number of the most abundant phytoplankton species cells (ind./L) from different depth in Lake Durowskie from 26th June to 30th June 2017 (Inf – inflow; A2 – Aerator 2; Mid. 2 – Middle 2; Mid.1 – Middle 1; Outf – outflow; B1 – Beach 1; B2 – Beach 2).

Depth	0m	0-3m	0-3m	0-3m	0-3m	0-1m	0m	0m
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2
Cyanoprokaryota - cyanobacteria								

<i>Anabaena lemmermanii</i> P. Richter	3200
<i>Anabaena flos-aquae</i> Brebisson	35200
<i>Aphanizomenon flos-aquae</i> (L.) Ralfs	11200	1200	3733.3
<i>Aphanocapsa incerta</i> (Lemm.) Cronberg et Komarek	46400	3600	800	4000	4533	12800	.	4000
<i>Chroococcus turgidus</i> (Kütz.) Naeg.	800	.
<i>Chroococcus limneticus</i> Lemm.	.	2400
<i>Limnothrix redekei</i> (Van Goor) Meffert	31200	3200	21200	1200	2800	800	.	400
<i>Microcystis aeruginosa</i> Kützing	1600	800
<i>Merismopedia punctata</i> Meyen	800	.
<i>Planktolyngbya limnetica</i> (Lemm.) Kom. – Legn. Et Cronenberg	.	11200
<i>Planktothrix agardhii</i> (D.C. ex Gom.) Anagn. et Kom.	112000	2000	6000	800	800	.	.	.
<i>Pseudanabaena limnetica</i> (Lemm.) Kom.	.	800	800
<i>Woronichina naegeliana</i> (Unger) Elenkin	800
Total	241600	25200	31733	6000	8133	13600	1600	4400
Bacillariophyceae- diatoms								
<i>Amphora ovalis</i> Kützing	4800
<i>Achnanthes minutissima</i> Kützing
<i>Asterionella formosa</i> Hasall	40000
<i>Cocconeis placentula</i> Ehr.	2400
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2
<i>Cocconeis pediculus</i> Ehr.	1600
<i>Cyclotella meneghiniana</i> Kützing	800
<i>Cyclotella ocellata</i> Pant.	8800	22933	4400	1600	8800	4000	.	.
<i>Cyclotella radiosa</i> (Grun.) Lemm.	26400	5600	3200	4000	5400	5200	.	.
<i>Cymbella amphycephala</i> Grun.	1600

<i>Cymbella lanceolata</i> (Ehr.) Kirchner	800
<i>Cymbella minuta</i> Hilse ex Rabenhorst	12000	1200	.	800	1600	800	800	.
<i>Cymatopleura solea</i> (Breb.) W. Smith	800
<i>Epithemia sorex</i> Kützing	800
<i>Fragilaria construens</i> (Ehr.) Grunow	47200
<i>Fragilaria crotonensis</i> Kitton	47200	3200	4000	.
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot	8000	4000	4400	1200	1000	2000	.	400
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen	1600
<i>Gomphonema olivaceum</i> (Horn.) Breb.	3200
<i>Hippodonta capitata</i> (Ehr.) L-Bert., Metz. & Witkowski
<i>Meridion circulare</i> (Grev.) C. Agardh	8800	.	800
<i>Navicula cincta</i> (Ehr.) Ralfs	3200
<i>Navicula lanceolata</i> (Agardh) Ehr.	800
<i>Navicula radiosa</i> Kützing	6400	800	800	.
<i>Navicula reinhardtii</i> Grun.	800
<i>Naviula tripunctata</i> (Muller) Bory	1600
<i>Nitzschia palea</i> (Kütz.) W. Smith	800
<i>Nitzschia recta</i> Hantzsch	800
<i>Nitzschia sigmoidea</i> (Ehr.) W. Smith	800
<i>Pinnularia viridis</i> (Nitzsch) Ehr.	800
<i>Rhopalodia gibba</i> (Ehr.) Muller	3200
<i>Stephanodiscus astrea</i> (Ehr.) Grunow	.	800
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2
Total	236000	38533	12800	7600	16800	12000	5600	400
Chlorophyta- green algae								
<i>Botryococcus braunii</i> Kutzing	16800	2133

<i>Closterium acutum</i> var. <i>variabile</i> (Lemm.) Krieg.	.	800
<i>Coelastrum astroideum</i> De Notaris	10400	2000	22133	1200	3200	7200	8800	19200
<i>Coelastrum microporum</i> Naegel.	1600	1600	.	800
<i>Coelastrum reticulatum</i> (Dang.) Senn	800	2133	800
<i>Cosmarium regnellii</i> Wille	800	.	800
<i>Crucigenia tetrapedia</i> (Kirch.) W. et G.S. West	.	.	2400
<i>Desmodesmus communis</i> (Hegew.) Hegew.	6400	1600	800	.
<i>Dictyosphaerium pulchellum</i> Wood	4800
<i>Elkatothrix gelatinosa</i> Wille	4800	800	.
<i>Eudorina elegans</i> Ehr.	.	.	.	800	.	.	4000	.
<i>Kirchneriella contorta</i> var. <i>elegans</i> (Schm.) Bohlin	3200	800
<i>Monoraphidium contortum</i> (Thur.) Kom.-Legn.	16800	2400	800
<i>Oocystis lacustris</i> Chodat	22400	4000	2133	1200	2400	2400	.	400
<i>Pediastrum boryanum</i> (Turpin) Meneg.	4000	2800	1867	1200	3200		3200	800
<i>Pediastrum duplex</i> Meyen	800	1600	800	800	2667	34400	800	.
<i>Phacotus lenticularis</i> (Ehr.) Stein	23200	12200	4400	20800	10400	6800	31200	11600
<i>Scenedesmus acuminatus</i> (Lager.) Chodat	2400	800
<i>Scenedesmus obtusus</i> Meyen
<i>Sphaerocystis planctonica</i> (Korsikov) Burrelly	338400	14000	118600	2987	41800	107200	187200	30133
<i>Staurastrum gracile</i> Ralfs	31200	1867	1600	.	.	.	800	.
<i>Tetraedron minimum</i> (A. Br.) Hansgirg	11200	1200
<i>Tetrastrum staurogeanieforme</i> (Schroed.) Lemm.	.	.	3200
<i>Tetrstrum komarekii</i> Hindak	2400	.	800	.	.	.	8000	.
Total	502400	51933	160333	56667	63667	158000	245600	62133
Cryptophyta- cryptophytes								
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2

<i>Cryptomonas erosa</i> Ehrenberg	31200	11467	4000	4000	9067	6000	4800	800
<i>Cryptomonas marssonii</i> Skuja	16000	.	800
<i>Cryptomonas ovata</i> Ehrenberg	3200	.
<i>Cryptomonas rostrata</i> Troitz. emend I. Kiselev	11200	1400	1200	800	2800	2000	800	1600
<i>Rhodomonas minuta</i> Skuja	44800	4533	1867	4000	1333	2400	3200	.
Total	103200	17400	7867	8800	13200	10400	12000	2400
<i>Dinophyta- dinophytes</i>								
<i>Ceratium hirundinella</i> (F. B. Müller) Bergh	9600	8800	2200	1600	5200	800	800	.
<i>Peridiniopsis berolinense</i> (Lemm.) Bourrelly	1600	.	3200	2400	2400	.	.	800
<i>Peridinium cinctum</i> (O.F. Müller) Ehrenberg	32000	11200	4400	1333	10133	1600	.	533
<i>Peridiniopsis cuningtonii</i> Lemm.	13600	2933	3400	2400	6400	2400	800	2133
<i>Peridiniopsis elpatiewskyi</i> (Ostenf.) Bourrelly	6400	1200	.	1600	1867	.	.	.
Total	63200	24133	13200	9333	26000	4800	1600	3467
<i>Euglenophyta- euglenoids</i>								
<i>Colacium vesiculosum</i> Ehr.	800	.	.
<i>Euglena acus</i> Ehr.	.	.	.	800
<i>Trachelomonas hispida</i> (Perty) Stein	1600	.	800
Total	1600	.	800
<i>Chrysophyceae- chrysophytes</i>								
<i>Dinobryon divergens</i> Imhof	16720000	87442	13800	.	.	.	36000	.
<i>Erkenia subaequiciliata</i> Skuja	114400	30200	2000	9867	14600	8400	4000	533
Total	16834400	117642	15800	9867	14600	8400	40000	533

ANNEX 4. Average biomass of the most abundant phytoplankton species (mg/L) from different depth in Lake Durowskie from 26th June to 30th June 2017 (Inf – inflow; A2 – Aerator 2; Mid. 2 – Middle 2; Mid.1 – Middle 1; Outf – outflow; B1 – Beach 1; B2 – Beach 2).

Depth	0-3m	0-3m	0-3m	0-3m	0-3m	0-3m	0-3m	0-3m
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2
Cyanoprokaryota - cyanobacteria								
<i>Anabaena flos-aquae</i> Brebisson	0.1090	-	-	-	-	-	-	-
<i>Anabaena lemmermanii</i> P. Richter	0.0060	-	-	-	-	-	-	-
<i>Aphanizomenon flos-aquae</i> (L.) Ralfs	0.0220	0.0020	0.0070	-	-	-	-	-
<i>Aphanocapsa incerta</i> (Lemm.) Cronberg et Komarek	0.0150	0.0144	0.0004	0.0010	0.0220	0.0040		0.0010
<i>Chroococcus turgidus</i> (Kütz.) Naeg.	-	-	-	-	-	-	0.0002	-
<i>Chroococcus limneticus</i> Lemm.	-	0.0002	-	-	-	-	-	-
<i>Limnithrix redekei</i> (Van Goor) Meffert	0.0090	0.0021	0.0063	0.00015	0.0009	0.0012		0.0010
<i>Merismopedia punctata</i> Meyen	-	-	-	-	-	-	0.0001	-
<i>Microcystis aeruginosa</i> Kützing	0.0010	0.0004						
<i>Planktolyngbya limnetica</i> (Lemm.) Kom. – Legn. Et Cronenberg	-	0.0040	-	-	-	-	-	0.0010
<i>Planktothrix agardhii</i> (D.C. ex Gom.) Anagn. et Kom.	0.1410	0.0025	0.0075	0.0010	0.0010	-	-	-
<i>Pseudanabaena limnetica</i> (Lemm.) Kom.	-	0.0003	-	-	-	0.0010	-	0.0010
<i>Woronichina naegeliana</i> (Unger) Elenkin	0.0010	-	-	-	-	-	-	-
Total	0.0380	0.0032	0.0053	0.0007	0.0081	0.0018	0.0002	0.0010
Bacillariophyceae - diatoms								
<i>Achnanthes minutissima</i> Kützing	-	-	-	-	0.0011	-	-	-
<i>Amphora ovalis</i> Kützing	0.0240	-	-	-	-	0.0040		0.0090
<i>Asterionella formosa</i> Hasall	0.0070	-	-	-	-	-	-	-
<i>Cocconeis pediculus</i> Ehr.	0.0110	-	-	-	-	-	-	-
<i>Cocconeis placentula</i> Ehr.	0.0160	-	-	-	-	-	-	-
<i>Cyclotella meneghiniana</i> Kützing	0.0010	-	-	-	-	-	-	-

Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2
<i>Cyclotella ocellata</i> Pant.	0.0090	0.0240	0.0040	0.0020	0.0090	0.0040	-	-
<i>Cyclotella radiosa</i> (Grun.) Lemm.	0.0330	0.0067	0.0040	0.0050	0.0068	0.0065	-	-
<i>Cymatopleura solea</i> (Breb.) W. Smith	0.0200	-	-	-	-	-	-	-
<i>Cymbella amphicephala</i> Grun.	0.0090	-	-	-	-	-	-	-
<i>Cymbella lanceolata</i> (Ehr.) Kirchner	0.0080	-	-	-	-	-	-	-
<i>Cymbella minuta</i> Hilse ex Rabenhorst	0.0220	0.0020		0.0010	0.0030	0.0010	0.0010	
<i>Epithemia sorex</i> Kützing	0.0010	-	-	-	-	-	-	-
<i>Fragilaria construens</i> (Ehr.) Grunow	0.0180	-	-	-	-	-	-	-
<i>Fragilaria crotonensis</i> Kitton	0.0010	0.0010			0.0070	0.0020	0.0020	
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot	0.0110	0.0030	0.0060	0.0015	0.0035	0.0025		0.0010
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen	0.0040	-	-	-	-	0.0010	-	-
<i>Gomphonema olivaceum</i> (Horn.) Breb.	0.0020	-	-	-	-		-	-
<i>Hippodonta capitata</i> (Ehr.) L-Bert., Metz. & Witk.	-	-	-	-	-	0.0010	-	-
<i>Meridion circulare</i> (Grev.) C. Agardh	0.0140	-	0.0010	-	-		-	-
<i>Navicula cincta</i> (Ehr.) Ralfs	0.0020	-	-	-	-	0.0010	-	-
<i>Navicula lanceolata</i> (Agardh) Ehr.	0.0040	-	-	-	-		-	-
<i>Navicula radiosa</i> Kützing	0.0160	0.0020	-	-	0.0020	0.0030	0.0020	0.0040
<i>Navicula reinhardtii</i> Grun.	0.0050	-	-	-	-	-	-	-
<i>Naviula tripunctata</i> (Muller) Bory	0.0040	-	-	-	-	0.0020	-	-
<i>Nitzschia palea</i> (Kütz.) W. Smith	0.0010	-	-	-	-	-	-	-
<i>Nitzschia recta</i> Hantzsch	0.0080	-	-	-	-	-	-	-
<i>Nitzschia sigmoidea</i> (Ehr.) W. Smith	0.0090	-	-	-	-	-	-	-
<i>Pinnularia viridis</i> (Nitzsch) Ehr.	0.0060	-	-	-	-	-	-	-
<i>Rhopalodia gibba</i> (Ehr.) Muller	0.0160	-	-	-	-	-	-	-
<i>Stephanodiscus astrea</i> (Ehr.) Grunow		0.0060		-	-	-	-	-
Total	0.0101	0.0063	0.0038	0.00238	0.0046	0.0025	0.0017	0.0047
Chlorophyta - green algae								

<i>Botryococcus braunii</i> Kutzing	0.0170	0.0011	-	-	0.0090	0.0020	-	-
<i>Cladophora glomerata</i> (L.) Kutzing	-	-	-	-	-	-	-	0.0030
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2
<i>Closterium acutum</i> var. <i>variabile</i> (Lemm.) Krieg.	-	0.0004	-	-	-	-	-	-
<i>Coelastrum astroideum</i> De Notaris	0.0330	0.00675	0.0707	0.0040	0.0105	0.0230	0.0280	0.1240
<i>Coelastrum microporum</i> Naegel.	0.0050	0.0055	-	0.0030	-	-	-	-
<i>Coelastrum reticulatum</i> (Dang.) Senn	0.0030	0.0073	0.0030	-	0.0080	0.0030	-	-
<i>Cosmarium regnellii</i> Wille	0.0001	-	0.0001	-	-	-	-	-
<i>Crucigenia tetrapedia</i> (Kirch.) W. et G.S. West	-	-	0.0001	-	-	-	-	-
<i>Desmodesmus communis</i> (Hegew.) Hegew.	0.0060	0.0010	-	-	0.0007	-	0.0010	-
<i>Dictyosphaerium pulchellum</i> Wood	0.0230	-	-	-	-	-	-	-
<i>Elkatothrix gelatinosa</i> Wille	0.0050	-	-	-	0.0030	0.0020	0.0010	-
<i>Eudorina elegans</i> Ehr.	-	-	-	0.0220	-	-	0.0700	-
<i>Eutetramorus planctonicus</i> (Kors.) Bourr.	-	0.0550	-	-	-	-	-	-
<i>Kirchneriella contorta</i> var. <i>elegans</i> (Schmidle) Bohlin	0.0003	0.0006	-	-	0.0002	-	-	-
<i>Monoraphidium contortum</i> (Thur.) Kom.-Legn.	0.0030	0.0040	0.0001	-	0.0008	-	-	-
<i>Oocystis lacustris</i> Chodat	0.0570	0.0100	0.0053	0.003	0.006	0.0060	-	0.0020
<i>Pediastrum boryanum</i> (Turpin) Meneg.	0.1180	0.0828	0.0547	0.0355	0.0943	-	0.0940	0.0470
<i>Pediastrum duplex</i> Meyen	0.0230	0.0470	0.0230	0.0280	0.0853	1.0110	0.0240	-
<i>Phacotus lenticularis</i> (Ehr.) Stein	0.0180	0.0097	0.0035	0.0165	0.0083	0.0055	0.0250	0.0190
<i>Scenedesmus acuminatus</i> (Lager.) Chodat	0.0020	0.0006	-	-	0.001	-	-	-
<i>Scenedesmus obtusus</i> Meyen	-	-	-	-	-	-	-	0.0010
<i>Sphaerocystis planctonica</i> (Korsikov) Bourrelly	0.772	0.0313	0.26975	0.0677	0.0955	0.2445	0.4270	0.2060
<i>Staurastrum gracile</i> Ralfs	0.0800	0.0047	0.0040	-	0.0060	0.0020	0.0020	-
<i>Tetraedron minimum</i> (A. Br.) Hansgirg	0.0010	0.00015	-	-	0.0004	-	-	-
<i>Tetrastrum staurogeanieforme</i> (Schroed.) Lemm.	0.0004	-	0.0001	-	0.0001	0.0020	0.0010	-
<i>Tetrstrum komarekii</i> Hindak	-	-	0.0003	-	-	-	-	-
Total	0.0614	0.0157	0.0334	0.0225	0.0206	0.1301	0.0673	0.0574

<i>Cryptophyta - cryptophytes</i>								
<i>Cryptomonas erosa</i> Ehrenberg	0.0510	0.0187	0.0063	0.0060	0.0147	0.0095	0.0080	0.0010
<i>Cryptomonas marssonii</i> Skuja	0.0200	-	0.0010		0.0010	-	-	0.0010
Site	Inf	A2	Mid.1	Mid.2	A1	Outf	B1	B2
<i>Cryptomonas ovata</i> Ehrenberg	-	-	-	-	-	-	0.0030	-
<i>Cryptomonas rostrata</i> Troitzskaja emend I. Kiselev	0.0240	0.0030	0.0020	0.0020	0.0060	0.0045	0.0020	0.0030
<i>Rhodomonas minuta</i> Skuja	0.0320	0.0033	0.0010	0.0030	0.0009	0.002	0.0020	-
Total	0.0317	0.0083	0.0026	0.0037	0.0056	0.0053	0.0037	0.0017
<i>Dinophyta - dinophytes</i>								
<i>Ceratium hirundinella</i> (F. B. Müller) Bergh	0.9290	0.8510	0.2123	0.1547	0.5030	0.0707	0.0770	-
<i>Peridiniopsis cuningtonii</i> Lemm.	0.0510	0.0340	0.0393	0.0278	0.074	0.0280	0.0090	0.0740
<i>Peridinium cinctum</i> (O.F. Müller) Ehrenberg	0.8490	0.2973	0.1163	0.0350	0.269	0.0420	-	0.0430
<i>Peridinopsis berolinense</i> (Lemm.) Bourrelly	0.0150	-	0.0285	0.0170	0.0200	-	-	0.0150
<i>Peridinopsis elpatiewskyi</i> (Ostenf.) Bourrelly	0.0510	0.0095	-	0.0130	0.0150	-	-	-
Total	0.3790	0.2979	0.0991	0.0495	0.1762	0.0490	0.0430	0.0440
<i>Euglenophyta - euglenoids</i>								
<i>Colacium vesiculosum</i> Ehr.	-	-	-	-	-	0.0020	-	-
<i>Euglena acus</i> Ehr.	-	-	-	-	0.0060	-	-	-
<i>Trachelomonas hispida</i> (Perty) Stein	0.0020	-	0.0010	-	0.0059	-	-	-
Total	0.0020	-	0.0010	-	0.0059	0.0020	-	-
<i>Chrysophyceae - chrysophytes</i>								
<i>Dinobryon divergens</i> Imhof	7.6240	0.0375	0.0060	-	0.0188	0.0030	0.0160	0.0020
<i>Erkenia subaequiciliata</i> Skuja	0.0060	0.0016	0.0003	0.0012	0.00075	0.0005	0.0002	0.0001
Total	3.815	0.0196	0.00313	0.0012	0.0098	0.0017	0.0081	0.0010

ANNEX 5. Comparison of periphyton species composition in different investigated sites from 26th June to 30st June 2017 in Lake Durowskie.

Diatom taxa	Site												p H	O	T	
	1	2	3	4	5	6	7	8	9	10	11	12				
<i>Achnanthes conspicua</i> Mayer							+									
<i>Achnanthes exigua</i> Grun.		+	+	+	+	+	+	+	+	+	+	+	+	4	1	7
<i>Achnanthes hungarica</i> (Grunow) Grun. in Cleve				+								+	4	4	6	
<i>Achnanthes lanceolata</i> (Breb.) Grunow	+	+	+	+		+		+				+	4	3	5	
<i>Achnanthes lanceolata</i> v. <i>elliptica</i> Cleve sensu Straub		+				+	+	+					4	-	-	
<i>Achnanthes lanceolata</i> var <i>lineate</i> Hustedt						+							-	-	-	
<i>Achnanthes minutissima</i> Kützing	+	+	+	+	+	+	+	+	+	+	+	+	3	1	7	
<i>Achnanthes minutissima</i> var. <i>affinis</i> (Grun.) Lange-Bertalot						+	+	+		+		+	4	-	-	
<i>Achnanthes minutissima</i> var. <i>gracillima</i> (Meister) Lange-Bertalot						+		+					4	-	1	
<i>Achnanthes rostrata</i> Lange-Bertalot				+									-	-	-	

<i>Amphipleura pellucida</i> (Kützing) Kützing		+											4	2	2
<i>Amphora ovalis</i> Kützing	+	+	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Amphora pediculus</i> (Kütz.) Grunow	+	+	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Asterionella formosa</i> Hass	+	+			+				+	+	+		4	2	5
<i>Aulacoseira granulate</i> (Ehr.) Simonsen		+							+				4	3	5
	1	2	3	4	5	6	7	8	9	10	11	12	p	O	T
													H		
<i>Caloneis bacillum</i> (Grun.) Meresz.	+	+	+	+		+		+				+	4	2	4
<i>Caloneissilicula</i> (Ehr.) Cleve		+											4	2	4
<i>Cocconeis pediculus</i> Ehr.	+	+	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Cocconeis placentula</i> Ehr.	+	+	+	+	+		+	+	+	+	+	+	4	3	5
<i>Cocconeis placentula</i> var. <i>linearis</i> Ehr.		+	+	+		+							-	-	-
<i>Cocconeis placentula</i> var. <i>lineate</i> Ehr.			+	+		+		+					4	3	5
<i>Cocconeis placentula</i> var. <i>pseudolineata</i> Geitler	+					+	+	+					-	-	-
<i>Craticula cuspidate</i> (Kützing) Mann W Round	+												4	3	5
<i>Cyclotella meneghiniana</i> Kütz.			+	+	+	+	+	+	+	+	+		4	5	5
<i>Cyclotella ocellata</i> Pant.	+	+	+	+	+	+	+	+	+	+	+	+	4	1	4
<i>Cyclotella operculata</i> (Ag.) Kützing		+	+		+		+		+	+	+	+	-	-	-
<i>Cyclotella radiosa</i> (Grun.) Lemm.	+	+	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Cymatopleura solea</i> (Breb.) W. Smith					+	+			+	+	+		4	3	5

<i>Cymbella affinis</i> Kützing	+		+	+	+	+	+	+	+	+	+	+	+	4	1	5
<i>Cymbella caespitosa</i> (Kützing) Brun.												+		0	-	7
<i>Cymbella cistula</i> (Ehr.) Kirchner	+	+	+	+	+				+		+			4	2	5
<i>Cymbella lanceolata</i> (Ehr.) Kirchner					+				+	+	+			4	1	7
<i>Cymbella microcephala</i> Grun.	+	+	+	+	+	+	+	+	+	+	+	+	+	4	1	4
<i>Cymbella minuta</i> Hilse ex Rabenhorst	+	+	+	+	+	+	+	+	+	+	+	+	+	3	-	-
<i>Cymbella prostrata</i> (Berkeley) Cleve					+	+	+		+	+	+			4	1	5
	1	2	3	4	5	6	7	8	9	10	11	12	p H	O	T	
<i>Cymbella tumida</i> (Bréb.) Van Heurck	+	+	+						+	+				4	1	4
<i>Cymbella turgida</i> (Greg.) Cleve	+													-	-	-
<i>Diatoma tenuis</i> Agardh					+					+	+			4	3	5
<i>Diatoma vulgare</i> Bory	+			+	+		+	+	+	+	+	+	+	5	2	4
<i>Diatoma vulgare</i> Bory Morphotyp ovalis		+				+								-	-	-
<i>Diploneis elliptica</i> (Kützing) Cleve					+				+	+	+			4	1	3
<i>Eunotia bilunaris</i> Ehr.							+	+						-	-	-
<i>Eunotia exigua</i> (Breb.) Rabenh.	+				+				+	+				1	2	7
<i>Eunotia faba</i> (Ehr.) Grun.					+				+		+			2	1	2
<i>Eunotia intermedia</i> (Krasske) Noerpel & Lange - Bertalot														2	-	1
<i>Eunotia praerupta</i> Ehr.	+	+	+		+	+	+	+	+	+	+	+	+	2	1	2

<i>Eunotia tenella</i> (Grun.) Hustedt													+	2	1	1
<i>Fragilaria capitata</i> Ehr.		+	+	+								+		-	-	-
<i>Fragilaria capucina</i> (Desm.) Rabenhorst	+		+	+	+	+	+	+	+	+	+	+	+	3	-	3
<i>Fragilaria constricta</i> Ehr.														2	-	1
<i>Fragilaria construens</i> (Ehr.) Grun.			+		+		+		+					4	1	4
<i>Fragilaria crotonensis</i> Kitton	+	+	+	+	+	+	+	+	+	+	+	+	+	4	2	3
<i>Fragilaria martyi</i> (Heribaud) Lange-Bertalot		+			+				+	+	+			-	-	-
<i>Fragilaria pinnata</i> Ehr.	+	+	+	+	+	+	+	+	+	+	+	+	+	4	1	7
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot	+	+	+	+		+	+	+	+	+	+	+	+	4	3	7
	1	2	3	4	5	6	7	8	9	10	11	12	p	O	T	
													H			
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen	+	+		+	+	+			+	+				4	2	7
<i>Fragilaria vaucheriae</i> (Kütz.) Carlson												+		-	-	-
<i>Gomphonema acuminatum</i> Ehr.	+				+	+		+	+	+	+	+	+	4	2	5
<i>Gomphonema angustatum</i> (Kütz.) Rabenhorst										+		+		-	-	-
<i>Gomphonema augur</i> Ehr.	+													4	1	4
<i>Gomphonema gracile</i> Ehr.	+	+			+				+	+	+			3	1	3
<i>Gomphonema intricatum</i> Kützing					+				+		+	+		-	-	-
<i>Gomphonema micropus</i> Kütz.					+				+	+	+			4	2	5
<i>Gomphonema olivaceoides</i> Hustedt	+	+			+	+			+	+	+	+		-	-	-

<i>Gomphonema olivaceum</i> (Horn.) Breb.	+	+	+	+	+	+	+	+	+	+	+	+	5	2	5
<i>Gomphonema parvulum</i> (Kütz.) Kütz.	+	+			+		+		+	+	+	+	3	4	5
<i>Gomphonema truncatum</i> Ehrenberg		+											4	2	4
<i>Hantzschia amphioxys</i> (Ehr.) Grunow					+		+		+		+		3	2	7
<i>Mastogloia smithii</i> Thwaites						+		+					4	-	-
<i>Meridion circulare</i> Ag.		+	+	+		+	+	+		+		+	4	2	7
<i>Navicula agrestis</i> Hustedt					+		+						3	-	-
<i>Navicula capitata</i> Patrick in Patrick & Reimer		+	+	+	+	+	+	+	+	+	+	+	4	3	4
<i>Navicula cincta</i> (Ehr.) Ralfs	+		+	+	+	+		+	+	+	+	+	4	3	5
<i>Navicula cryptocephala</i> Kütz.	+				+	+		+	+	+	+		3	3	7
<i>Navicula dicephala</i> (Ehr.) W. Sm.	+												-	-	-
	1	2	3	4	5	6	7	8	9	10	11	12	p	O	T
													H		
<i>Navicula gastrum</i> (Ehrenberg) Kützing											+		4	2	5
<i>Navicula gregaria</i> Donkin						+	+	+					4	4	5
<i>Navicula oblonga</i> Kützing											+		4	2	5
<i>Navicula placentula</i> (<i>Placoneis</i>) (Ehr.) Grunow					+				+	+	+		4	2	5
<i>Navicula radiosa</i> Kützing	+	+	+	+	+	+	+	+	+	+	+	+	3	2	4
<i>Navicula reinhardtii</i> Grun.			+	+			+			+			5	2	5
<i>Navicula tenella</i> Hustedt													-	-	-

<i>Navicula tripunctata</i> (O. F. Müller) Bory	+	+	+	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Navicula veneta</i> Kützing				+										4	4	5
<i>Navicula viridula</i> (Kütz.) Ehr.					+		+		+		+			4	2	5
<i>Nitzschia acicularis</i> (Kützing) W. Smith	+	+	+	+										4	4	5
<i>Nitzschia amphibian</i> Grunow	+	+		+	+				+	+	+	+		4	3	5
<i>Nitzschia incospicua</i> Grun.						+	+	+		+	+			4	3	5
<i>Nitzschia linearis</i> (Agardh) W. Smith												+		4	2	4
<i>Nitzschia micropus</i> (Kütz.)														-	-	-
<i>Nitzschia palea</i> (Kütz.) W. Sm.	+		+	+			+					+		3	4	6
<i>Nitzschia paleacea</i> Grun.			+	+		+	+	+				+		4	3	5
<i>Nitzschia recta</i> Hantzsch	+				+	+	+	+	+		+	+		4	2	7
<i>Nitzschia sigmaidea</i> (Ehr.) W. Sm.			+	+	+		+	+	+	+	+	+		4	3	5
<i>Pinnularia maior</i> (Kützing) Rabenhorst												+		3	2	4
	1	2	3	4	5	6	7	8	9	10	11	12	p H	O	T	
<i>Pinnularia viridis</i> (Nitzsch) Ehr.		+	+	+		+	+	+				+		3	3	7
<i>Rhoicosphaenia abbreviate</i> (Ag.) Lange-Bertalot		+										+		4	2	5
<i>Rhoicosphaenia curvata</i> (Kütz.) Grun.			+	+										-	-	-
<i>Rhopalodia gibba</i> (Ehr.) Müller	+	+	+			+			+	+				5	3	5
<i>Stauroneis phoenicentron</i> Ehr.	+		+	+		+		+						3	3	4

<i>Stephanodiscus astraea</i> (Ehr.) Grunow					+		+		+		+		-	-	-
<i>Stephanodiscus hantzschii</i> Grun.			+							+		+	5	4	6
<i>Surirella minuta</i> Breb.			+	+									4	3	5
<i>Surirella ovalis</i> Breb.					+				+		+		4	4	5
<i>Tabellaria fenestrata</i> (Lyngb.) Kützing	+	+				+	+	+		+			3	1	2