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ECOLOGICAL STATE OF LAKE DUROWSKIE BASED ON PHYTOPLANKTON AND PERIPHYTON IN JULY 2015

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Introduction

Algae, which is considered as the main primary producers and relevant source of oxygen for many water bodies, plays an important role in lake ecosystem. However, algal blooms, which may be caused by water pollution, have led to severe problems in eutrophic lakes, rivers and other water bodies over the worldwide, which may lead to oxygen depletion, fish gill clogging, production of toxicants and bad odours (Stumpf et al., 2009; Hudnell, 2010; Lewitus et al., 2012). Due to its sensitivity to environmental factors, phytoplankton has been widely used as indicator to evaluate changes in water bodies and has been used as a tool to monitor early warnings of water pollution (Thiébaut et al., 2006, Wu et al., 2012).

Wagrowiec is a town in northwestern Poland. The region around the town is rich in lakes and the town itself sits in the middle of Lake Durowskie. Lake Durowskie is a main attraction for tourists and locals. However it suffered from algal blooming problem since 1999, due to the direct disposal of sewage into it. Agricultural run-off, carrying loads of fertilizers, herbicides and pesticides, also brought additional nutrients such as phosphates and nitrates into the Lake. Due to the algal blooms, the transparency of the lake decreased and it turned out to be tons of dead fishes and other organisms. Consequently, the beaches area of the lake had to be closed and the tourism activities had to be stopped because of the really poor environmental conditions.

The local government of Wagrowiec paid much attention to the lake algal blooming problem. To change the situation, the government decided to start a restoration project for the Lake Durowskie in 2009. The important measures that have been taken so far include installation of aerators for the oxygenation of hypolimnion, addition of Iron sulphate (PIX) to sediment phosphorus and introduction of pike and pike-perch for bio-manipulation. Furthermore, the local government of Wagrowiec cooperated with Adam Mickiewicz University from Poznan to monitor and assess the effectiveness of restoration measures. The monitoring process is taking into consideration many aspects of lake quality assessment including hydrology, physic-chemical parameters, algae, macrophytes and micro-invertebrates communities etc.

From the reports of years before, it seems there is a good trend of the water quality of Lake Durowskie. This report will show and assess the present ecological state based on phytoplankton and periphyton, which was sampled in and around the shoreline of Lake Durowskie. The results will be compared with the data from last six years to assess the restoration project.

Material and Methods

Study area

Studies were conducted in Lake Durowskie, in the city of Wągrowiec between June 29 and July 03, 2015 in order to determine the ecological state of the lake.

Tab 1. Lake Morphometry

surface	143,7 ha
volume	11322900 m ³
max depth	14,6 m
mean depth	7,9 m
main tributary	Struga Gołaniecka
surface of the whole catchment area	236,1 km ²
surface of the direct catchment area	1581,3 ha
share of agricultural area	58,26 %
share of forest	33,52 %
urban area	8,25%

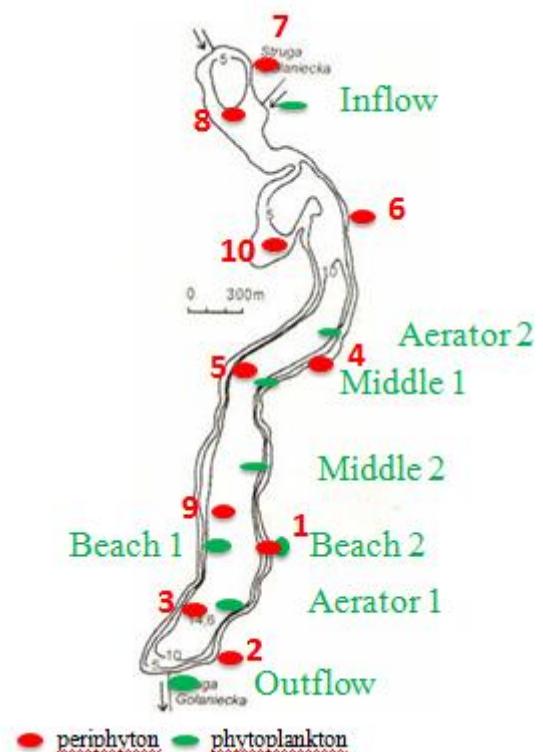


Fig 1. Sampling sites

Investigation were done on phytoplankton and periphyton. Water samples were collected from (8stations) on phytoplankton and (10 stations) on periphyton.

Methods

Phytoplankton

Phytoplankton samples were collected from the surface water at Inflow, Beach 1 and Beach 2, Outflow; and from 0, 1 and 2 meters depth in all Middle and Aerator stations. The samples were concentrated using a plankton net. Taxonomical analysis of sampling tok into consideration:

Number of phytoplankton species cells (ind./L)

We count species in 100 fields of view of the microscope. Then we count factual concentration:

sample concentration 30 ml from 30 l; $30\ 000\ \text{mm}^3 : 1.25\ \text{mm}^3 = 24\ 000$

24 000 – 301

x – 11

x = 800 – factual concentration

Both results multiply and get the individual number of cells per liter.

Biomass of phytoplankton species (mg/L)

To estimate the biovolume of 1 cell of particular algal species we were multiplied biovolume of 1 cell with the individual number of cells per liter. Next were put the comma on 9 place counting from the right side in direction left. The biomass was always given to 3 decimal places.

Indeces:

- **Mixed index of Nygaard**

Trophic status of the lake was estimated using the mixed index of Nygaard (Nygaard, 1949), following the equation:

$$\text{mixed index of Nygaard} = \frac{\text{Cyanobacteria} + \text{Chlorococcales} + \text{centric diatoms} + \text{euglenoids}}{\text{desmids}}$$

- **Jaccard Index**

The Jaccard Index was used to compare the similarity and diversity of the number of species between the years. We calculated it according to the formula (Jaccard, 1912):

$$J = \left(\frac{c}{a + b - c} \right) \times 100 \%,$$

Where J is index of Jaccard (percentage of species in common)

a – number of species in one year

b – number of species in another year

c – number of species in common

- **Shannon-Wiener and evenness indices**

In order to measure evenness and diversity of the species between different sites we used Shannon-Wiener and Evenness Indices according to Shaw (2003):

$$H' = - \sum_{i=1}^s p_i \log (p_i);$$

$$E = \frac{- \sum p_i \times \log (p)}{\log (S)};$$

Where H' is the Shannon index, p_i – percentage of individuals of certain species in the community; E – evenness (equitability); S – number of species in the community.

Periphyton

Periphyton species were collected from submerged stones in the 10 stations using a brush. The samples were conserved using Lugol solution and the species were identified in the laboratory.

Van Dam's ecological indicator values were used to estimate oxygen saturation, trophy and alkalinity in lake Durowskie (van Dam et al., 1994).

Diatom index

Diatom index is calculated only when in the sample you will find at least 10 species being sensitive to the trophy state (TJ_i).

- Trophy index (TJ)

$$TJ = \sum (TJ_i \cdot wTJ_i \cdot Li) / \sum (wTJ_i \cdot Li)$$

where:

TJ_i – value of the sensitivity of species for the trophy state;

w TJ_i - range of the tolerance of the algal species;

Li - number of specimens of the determined species divided by the number of all identified individuals in the sample

- Index of referential species (pGR)

$$pGR = (NB - (NC + ND)) / (NB + NC + ND)$$

where:

NB – number of referential species for all lakes;

NC – number of referential species for deep lakes and of degradation species in shallow lakes;

ND – number of degradation species for both kinds of lakes.

- Transformation of standardized value in the range from 1 (most beneficial) to 0 (least beneficial) according to equations:

$$Z - TJ = 1 - ((TJ - 1) \cdot 0.25)$$

$$Z - pGR = (pGR + 1) \cdot 0.5$$

Diatom index

$$DI = (Z-TJ + Z-pGR) / 2$$

and classification of the ecological state for Lake:

Ecological state	Value range
very good	>0,83
good	0,55
moderate	0,30
poor	0,15
bad	<0,15

Results and discussion

Phytoplankton

The total number of phytoplankton species collected in all the (8 station) was amount to 75 species and the total of dominated species was recorded in the Middle 2 and Inflow with moving gradually to other parts of the stations, as in Fig 2. This may be related to the high number of the diatom movement and current waves together with human activities which could result in rushing out the sediment phytoplankton around the lake to every stations recorded.

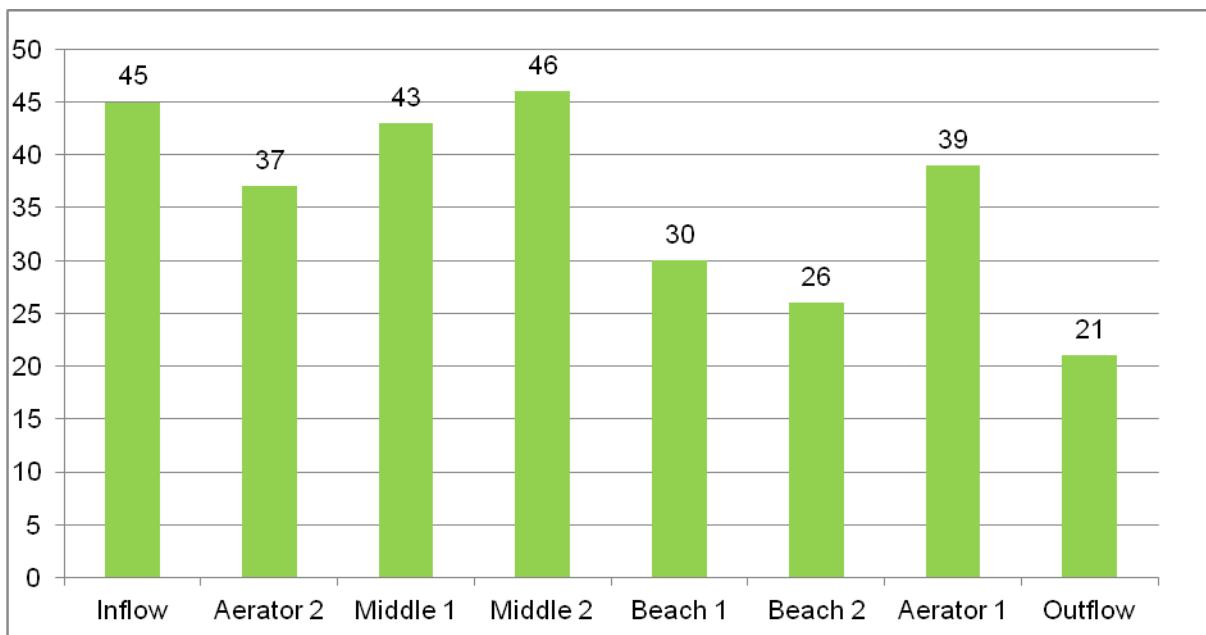


Fig 2. Total number of phytoplankton species from different sampling sites in 2015

Cyanobacteria were dominant and abundant species in the group of algae. Phosphorus and nitrogen flow with the Struga Gołaniecka stream to Lake Durowskie. The reason for the high abundance at the inflow is due to Cyanobacterias preference for nutrient rich waters. Morealso, the Fig 3. indicate hypertrophic state of the rivers water which was observed.

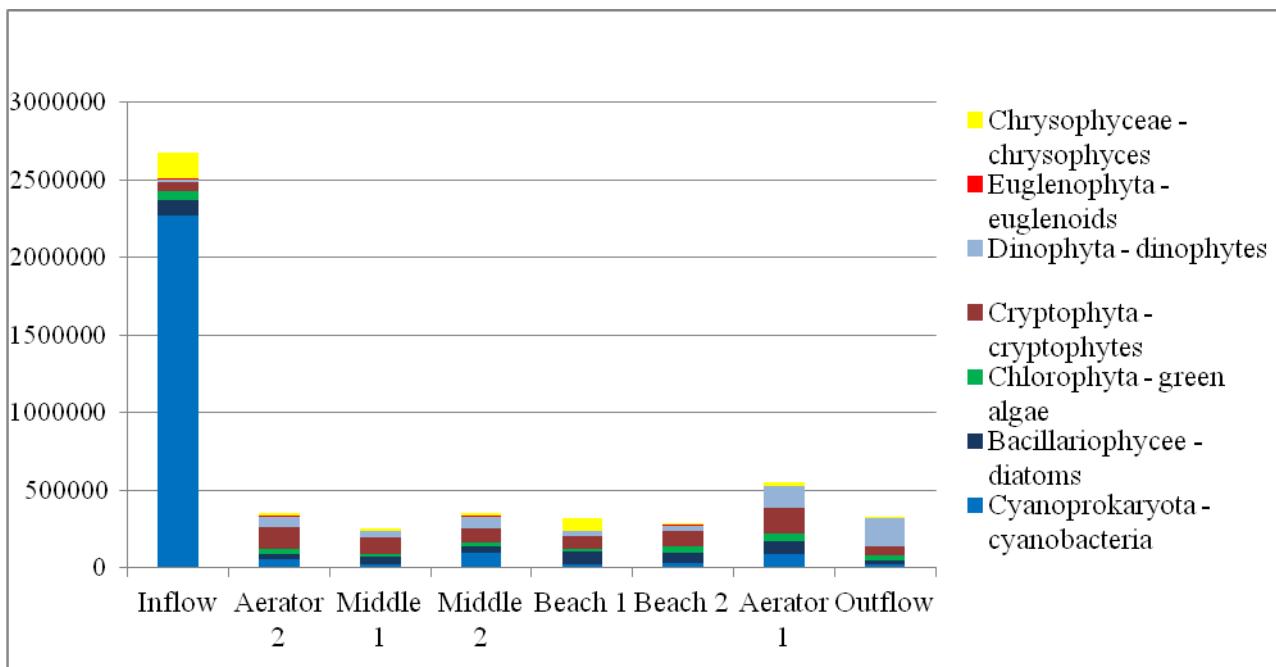


Fig 3. Abundance of each group of algae (number of cells)from different sampling sites in 2015

Figure 4., show how we calculate the biomass of every group of algae samples showing the result as dinophytes, contributed significantly to the total biomass of phytoplankton in different sites. It was related to its large and bigger cells of this group of Algae. Order group of algae shows low contribution and reduction in biomass due to small cells.

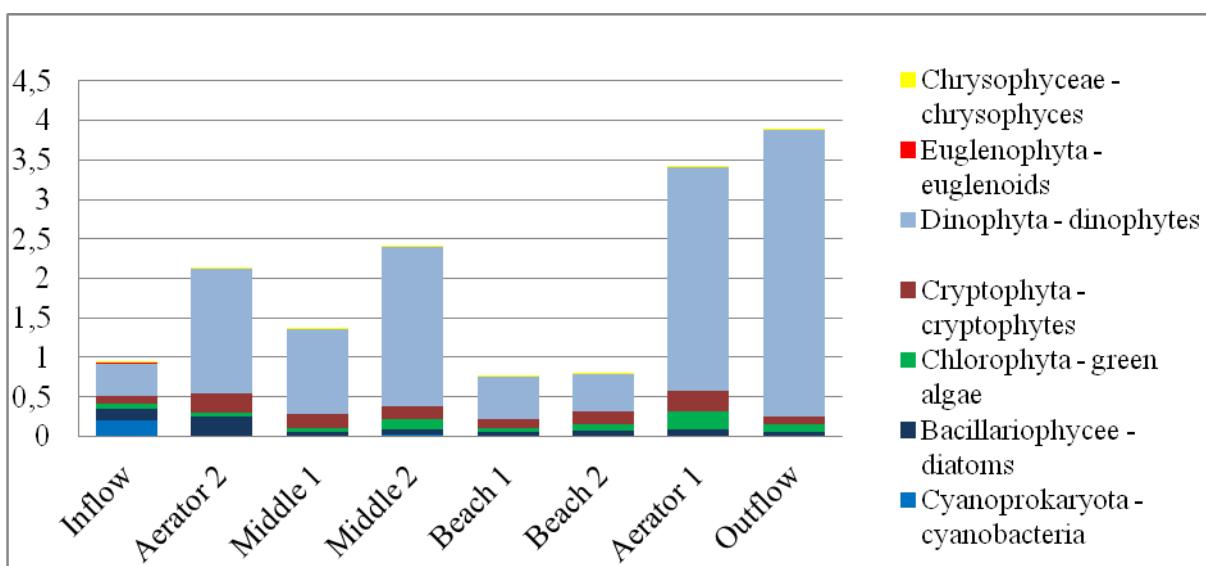


Fig 4.Biomass of each group of algae from different sampling sites in 2015

To the group of phytoplankton dominants of 17 species which were characterized by the greatest number of cells on individual sites were ranked. Species from cryptophytes (*Cryptomonas erosa*), dinoflagelates (*Peridiniopsis cunningtonii*, *Peridinium cinctum*), diatoms (*Fragilaria ulna*) and cyanobacterium (*Limnithrix redekei*) belonged to dominant taxa, which were found in a majority of sites (tab. 4). A large share of blue-green algae in the phytoplankton community connects with an activity of motor boats which create turbulence within a few metres depth taking water out from deeper layers to the surface layer (north sites A2 and M1 – dominance in the range 8-12%; south sites M2 and A1 - dominance in the range 10-27%). In the inflow of the lake only cyanobacteria were dominating (total 81%): *Limnothrix redekei* (64%), *Pseudanabaena limnetica* (10%) and *Aphanizomenon flos-aquae* (7%).

Table 4. Percentage share of dominating species in the total number of the phytoplankton on individual sites in Lake Durowskie (Infl – Inflow; A – aerator; M – Middle; Outfl – Outflow; B – Beach).

Mixed index of Nygaard (Nygaard, 1949), was using to determine the trophic state of the lake using the appropriate equation which is contained in the subsection Methods.

After assessing the samples base on the mixed index of Nygaard equation, we have 5 stations in hypertrophic state and 3 stations in eutrophic as show in the Table 2. It is more likely that this due to higher number of Cyanobacteria which are more abundant in the river that are flow into the open zone of lake. This still indicate better quality water, but we are hoping in future that we can have better improvement.

Tab 2. Trophic changes based on the mixed index of Nygaard over the years

Station	2008	2009	2010	2011	2012	2013	2014	2015
Inflow	-	-	1,8	17	9	19	3,8	17
Aerator 2	-	26	11,5	5	8	14	20	4,3
Middle 1	-	9	12,5	13	3	5,5	11	4,8
Middle 2	-	-	8,3	18	9	7,5	20	4
Beach 1	-	-	-	3	9	7	5	5,5
Beach 2	-	-	-	-	5	6	10	12
Aerator 1	9,7	16	8,3	9	7	8	9	6,7
Outflow	-	-	6,5	5	-	12	8	8

Legend	Result
Dystrophy	0 - 0,3
Oligotrophy	< 1,0
Mesotrophy	1,0 - 2,5
Eutrophy	3 - 5
Hypertrophy	5 - 43

We calculated the similarity and richness of the number of species between the years using the Jaccard index. In table 3 analyses indicates how many species in percentage are common in the previous years. According to this we can see that 52% of the species from 2014 while in 2015 we have 82 % which is very close to what we have in 2008. The similarities between 2008 and 2015 is large, however, most species of eutrophic appearing sporadically in the current year.

Tab 3.Jaccard index

YEAR	2009	2010	2011	2012	2013	2014	2015
2008	84	51	43	33	40	52	82
2009	-	48	28	20	29	35	39
2010	-	-	42	42	62	47	37
2011	-	-	-	34	58	47	50
2012	-	-	-	-	77	49	59
2013	-	-	-	-	-	52	78
2014	-	-	-	-	-	-	57

In 2015 there are 5 new species. However, we see in the graph Figu 5., that is two more species, resulting from the fact that the species also disappear.

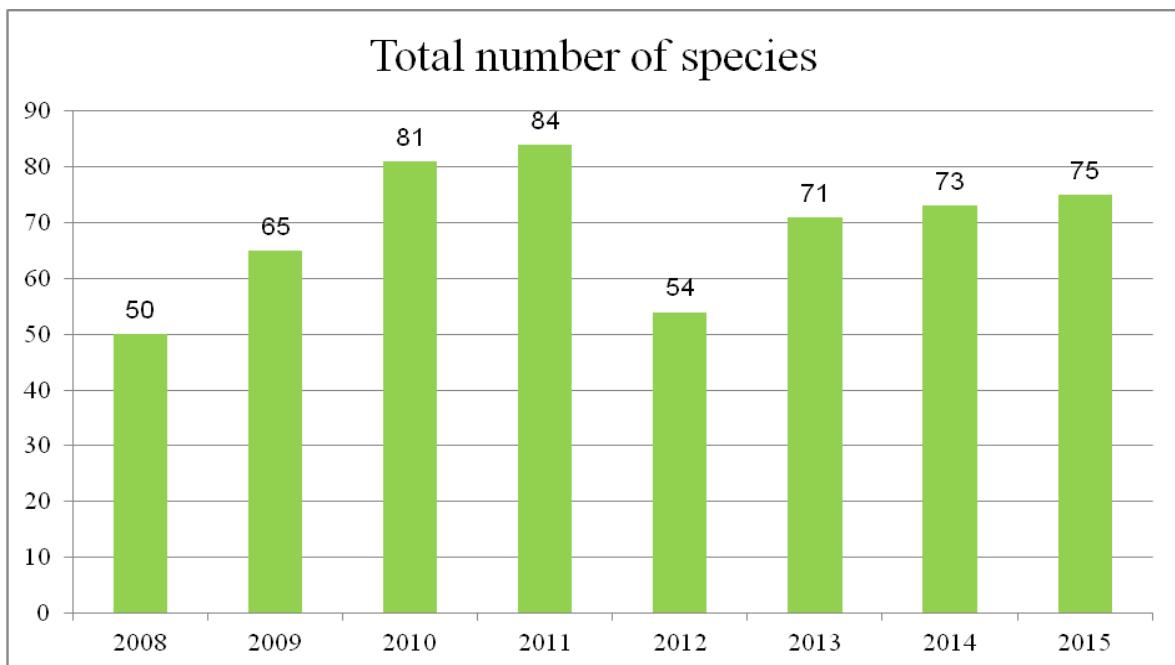


Fig 5. Total number of species over the years

In fig 6. highest diversity was noticed on station Inflow, which may be related to the flow from other parts of the river connected to the lake. The lowest Diversity Index was observed in station of Outflow which was connected to the current waves on the lake. The river (Struga Gołaniecka) provides additional species of benthos and periphyton, which detach from the surface by a current.

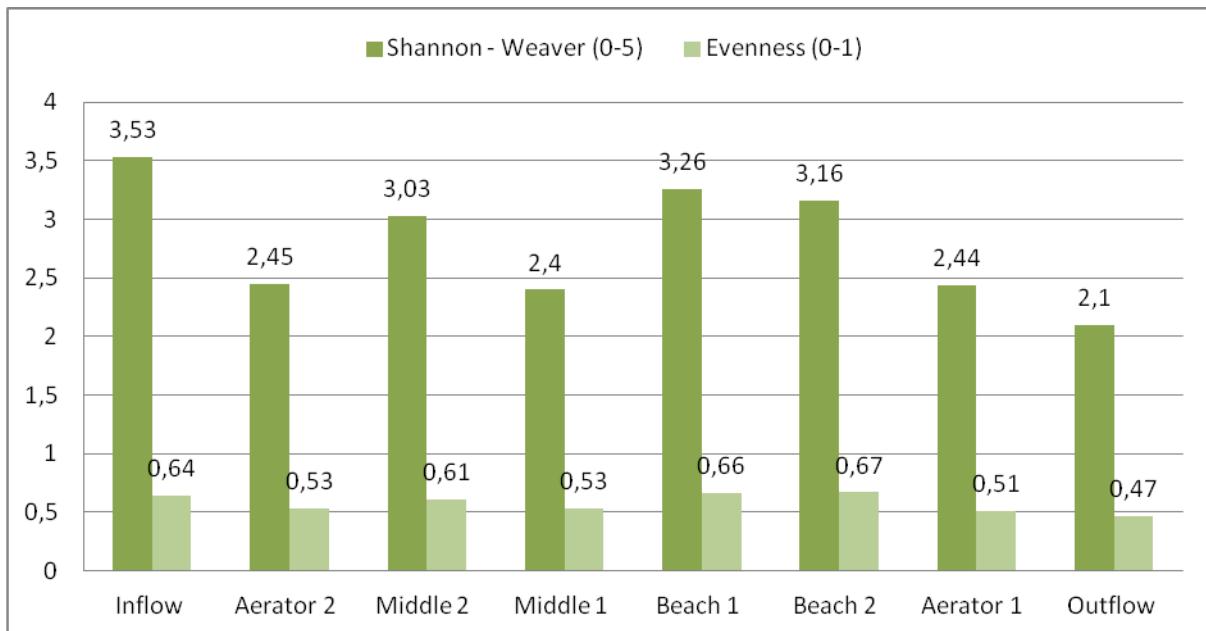


Fig 6. Evenness and Shannon-Weaver Index from different sampling sites in 2015

Biomanipulation

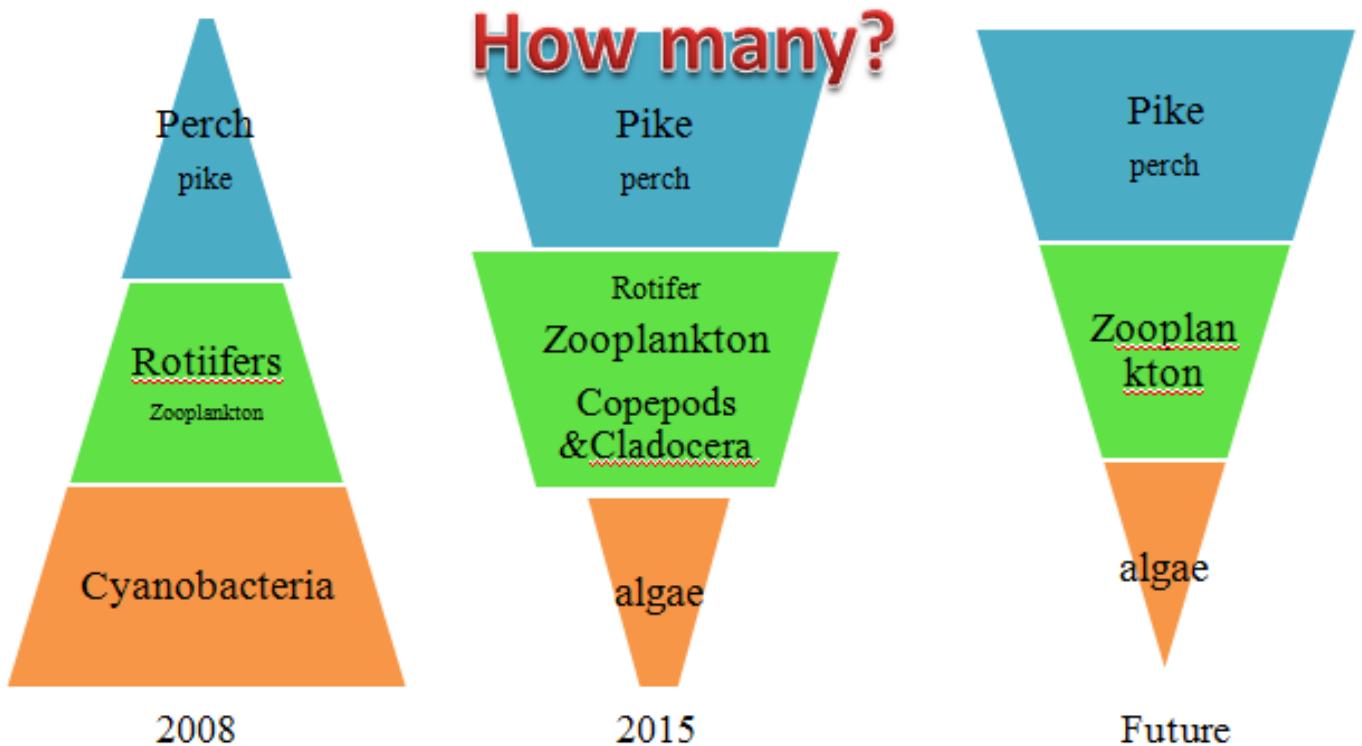


Fig 7. Grazing –trophic Pyramid- biomanipulation

Grazing –trophic Pyramid was used to assess the state of the lake due to its restoration, by introducing bio-manipulation to maintain the state of the lake. In comparing 2008 to 2015 the state of restoration through biomanipulation is working but still need close monitoring in-order for the Pike to be able to survive for a long period of time.

Periphyton

From all the ten sampling sites, 103 species of periphyton were found in Lake Durowskie. *Achnanthes minutissima* Kütz is the absolute dominant species as it appears in 9 sites out of 10 (tab. 5 and 6). Furthermore, it even shows as the 71% percentage of site 1. Compared to last year, the dominant species remain almost the same. However, they all appear in more sites than before. Besides *Achnanthes minutissima* Kütz, *Cymbella minuta* Hilse indicates increasing dominant status. This could be a good phenomenon as it prefers eutrophic water while the lake was hypertrophic before.

Table 5. Dominant species in periphyton community from different sampling sites in 2015.

Dominants in periphyton community	Species percentage from different sites (%)									
	1	2	3	4	5	6	7	8	9	10
<i>Achnanthes minutissima</i> Kütz	71	22	19	20	6	9	11		5	5
<i>Fragilaria crotonensis</i> Kitton	9						7	7	4	
<i>Cymbella affinis</i> Kütz.	8		5				5			11
<i>Cymbella minuta</i> Hilse	7	9			6	15	8		7	5
<i>Cocconeis placentula</i> Ehr.		11	12			7		11		
<i>Fragilaria pinnata</i>			6	6	5	7		5		
<i>Gomphonema olivaceum</i> (Horn.) Breb.				8	21				18	15

Table 6. Dominant species in periphyton community from different sampling sites in 2014.

Species	Site							
	1	2	3	4	5	6	7	8
<i>Achnanthes minutissima</i> Kütz	45.36	14.11	23.96	27.47		11.39	13.23	
<i>Achnanthes minutissima</i> var. <i>affinis</i> (Grun.) Lange-Bertalot						10.89		
<i>Amphora pediculus</i> (Kütz.) Grun.			20.22					
<i>Cymbella affinis</i> Kütz.		11.19						
<i>Cymbella minuta</i> Hilse						15.84		
<i>Cocconeis pediculus</i> Ehr.		10.67						
<i>Cocconeis placentula</i> Ehr.							16.52	
<i>Gomphonema olivaceum</i> (Horn.) Breb.				14.66	21.54			

For different species, they could represent for different trophic conditions. Information about some of the dominant species is listed following.

Achnanthes minutissima Kütz is a hypereutraphentic species, therefore it can easily adapt to any trophic conditions. This is probably the reason why it was abundant in the majority of the sites. However, it requires high oxygen saturation (>75%) and circumneutral pH (van Dam et al, 1994).

Gomphonema olivaceum (Horn.) Breb. exclusively occurs at pH >7, and prefers high oxygen saturation. *Gomphonema olivaceum* is a eutraphentic species (van Dam et al, 1994).

Cymbella affinis Kütz. is an alcaphilous and eutraphenic species, with a preference for waters highly saturated in oxygen.

Cymbella minuta Hilse occurs in lakes with neutral pH. It is a common species of Central Europe. *Cymbella minuta* is not a sensitive species to habitat alteration, as it also occurs in antropogenically disturbed areas.

Cocconeis placentula Ehr. requires only moderate levels of dissolved oxygen (>50%). It is eutraphentic and alcanophilous. *Cocconeis placentula* is the only dominating species at site 8, which is close to the inflow of the lake.

Preferences of periphyton

For different species, they have different preferences of living conditions, such as pH, dissolved oxygen, trophy state etc. Here in this report, we show the maps (Fig. 8.-10.) of the preferences of priphyton species about pH, oxygen and trophy state. From the maps, we could see more than 60% of the species prefer alkaline water; about half of the species prefer high oxygen concentration, some even prefer up to 100 percentage of dissolved oxygen; more than 50% species share the same trophy state preference as eutrophic (hypertrophic included).

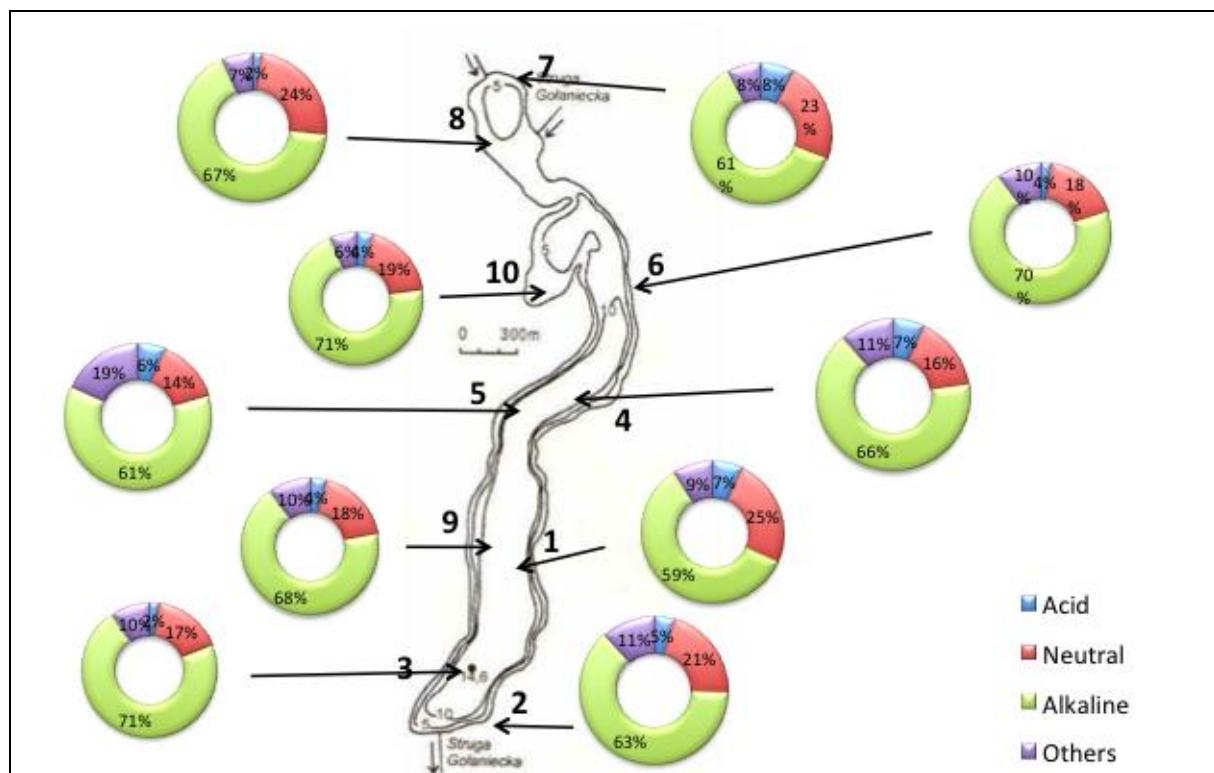


Fig 8. Preferential pH value of periphyton among all the ten sites in 2015.

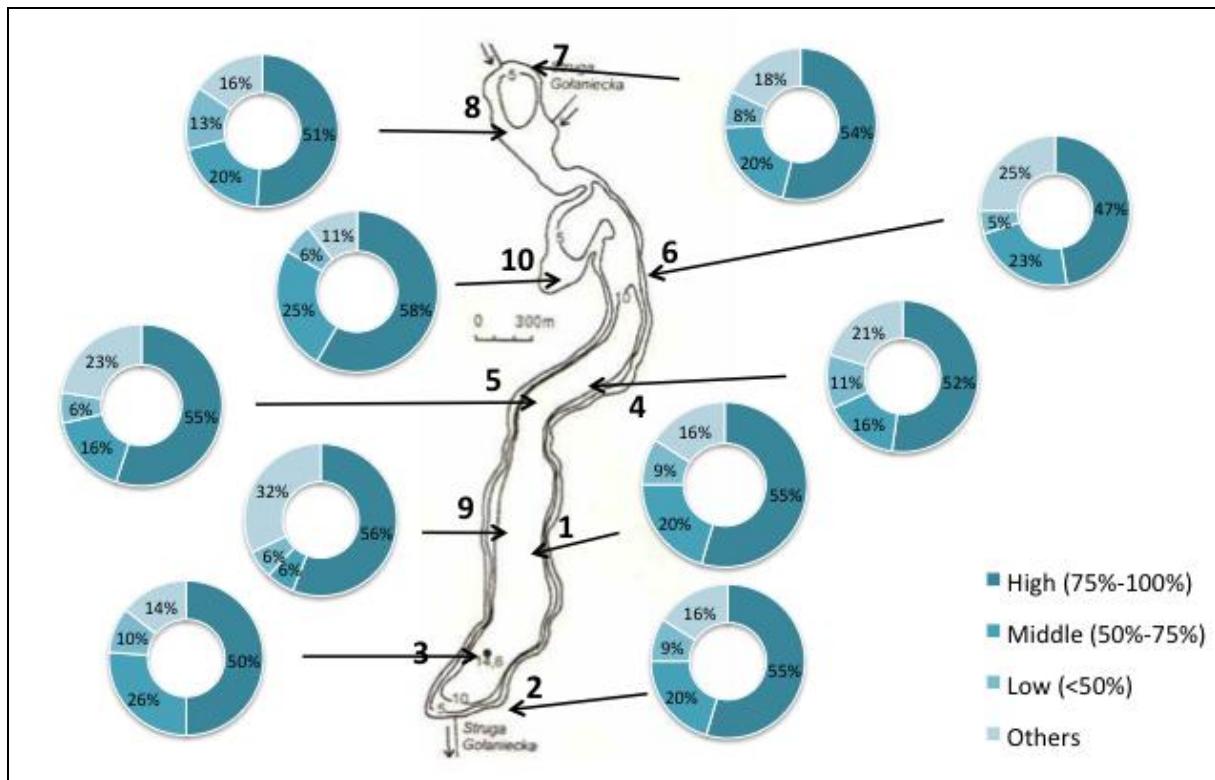


Fig 9. Preferential oxygen value of periphyton among all the ten sites in 2015.

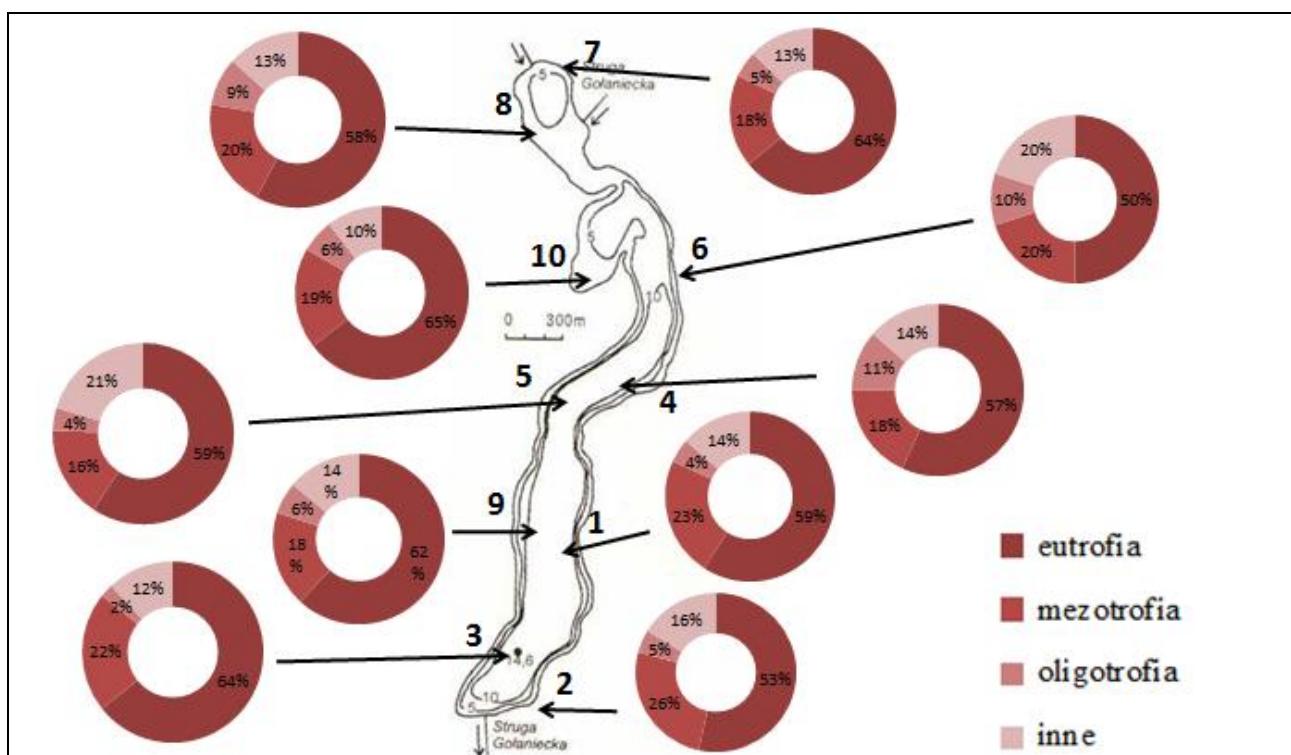


Fig 10. Preferential trophy state of periphyton among all the ten sites in 2015.

Diatom index

Diatoms are unicellular, photosynthetic, eukaryotic algae found throughout the world's oceans and freshwater systems (Armbrust et al., 2004). Many of the diatoms are sensitive to trophic state of water, therefore they can provide valuable information for monitoring rivers, particularly on organic pollution (Wu, 1999). To make the monitoring work more accurate, we sampled 10 sites this year, 2 more than the previous years.

As the same as previous years, the ecological state of Lake Durowskie varied from poor to moderate levels (Fig 11.). Only three sites are in moderate level and almost all the sites present a decreasing trend of the ecological state. The good point is that site 3 has a better situation of poor rather than bad in the year 2014. Broadly speaking, the lake ecological state remains stable.

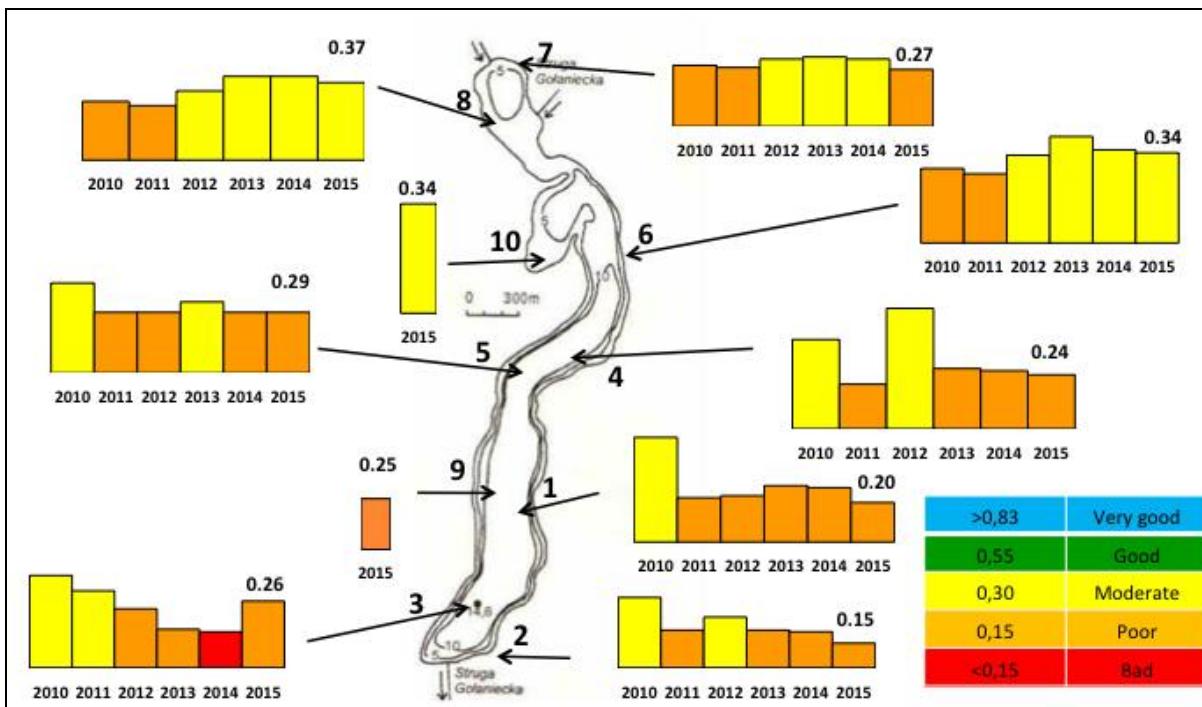


Fig. 11. Ecological state of the lake in different stations based on Diatom Index in 2015

We present here also the ecological state of Lake Durowskie in different geograph. The diatom indices are presented by the mean values for northern (site 4, 5, 6, 7, 8, 10) and southern (site 1, 2, 3, 9) parts. We could see that, the ecological state of the nouthern part is better than the southern part. Furthermore, there is an increasing trend, while the southern part remains the poor situation.

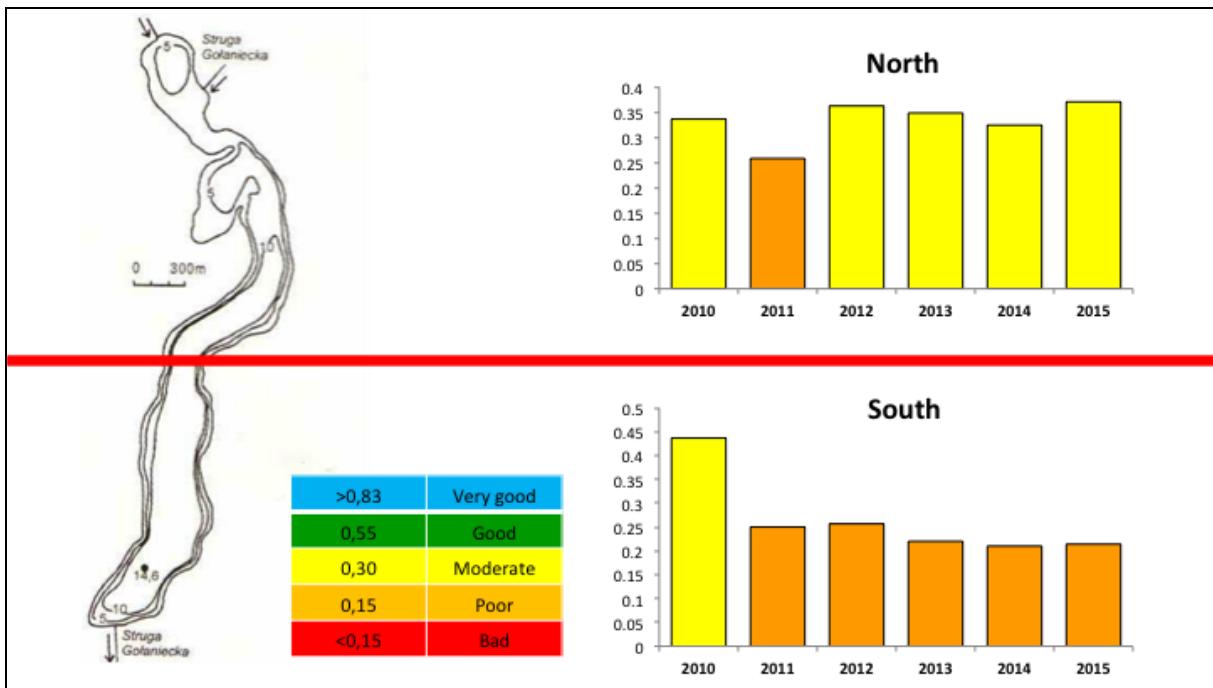


Fig. 12. Ecological state of Southern and Northern part of the lake based on Diatom Index in 2015

Red algae: *Hildenbrandia rivularis*

Hildenbrandia rivularis (fig. 13.) is a cosmopolitan red algae species, which prefers fresh flowing water habitats, but also occurs in standing water. It is considered to be a bioindicator of good water quality. However, it occurs right across the nutrient gradient, and the presence of stable substrata and clear water is probably more important than the nutrient concentration per se (Kelly&King, 2007).

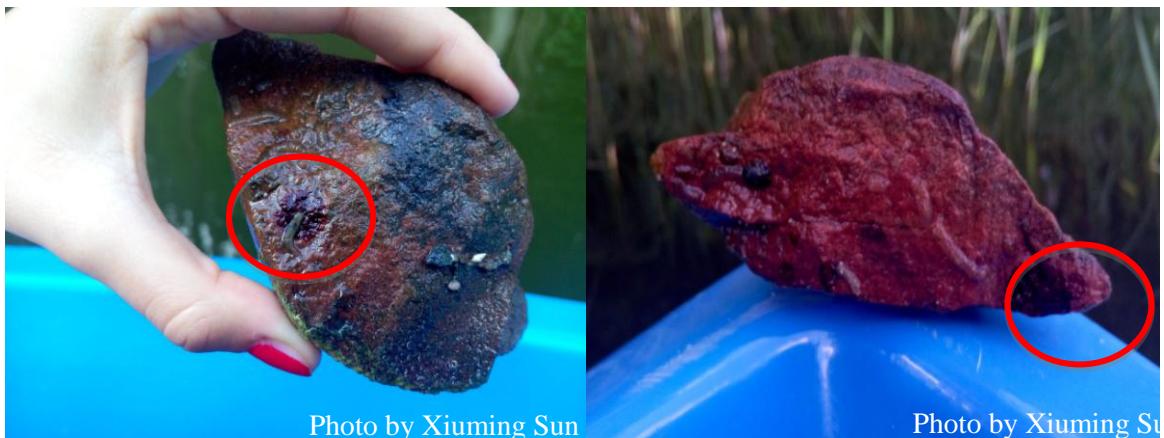


Fig. 13. Photos of red algae appeared on the stones in 2015.

The red algae first appeared in 2010, and an increasing abundance of these algae had been observed (fig 14.). This year the red algae appears at the same allocation as last year, along the south-east region. That could be because that, the dominant wind direction in this area is from the west, and this creates constant wave erosion and oxygenation of the water in the south-east region.

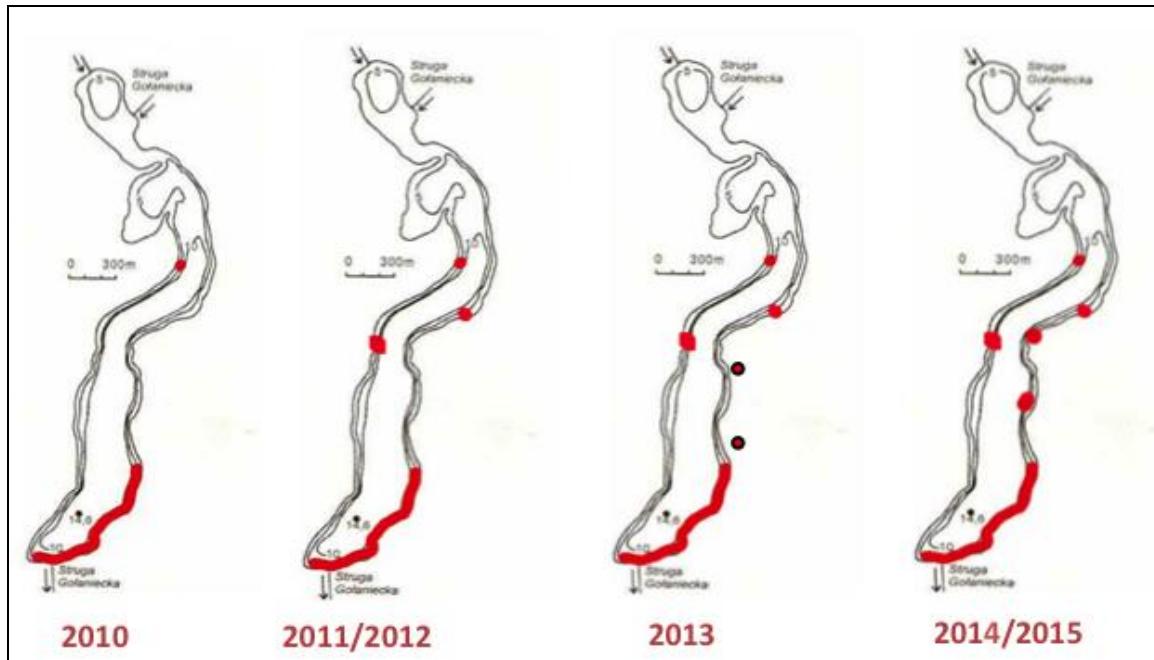


Fig 14. Changes in the abundance of *Hildebrandia rivularis*

Conclusions

From the analyses of our results, the ecological state of Lake Duroske is still eutrophic, and the hypertrophic situation in the inflow remains the same as before. We can only observe the high inflow of cyanobacteria but not inflow, which indicates that we should pay more attention to the upstream of the river and also the lakes up there.

In the research of this year, 5 new species of phytoplankton were found. This suggests the lake is restructuring into the better direction. In addition, the decrease of the phytoplankton density also turns out to be the consequence of bio-manipulation.

The southern part of the lake remains poor ecological state needs to be paid more attention. This could be due to higher anthropogenic pressure in the littoral zone, also could be due to the less distribution of macrophytes. Macrophytes can absorb nutrients to certain extent. From this point of view, it is necessary to protect the macrophytes along the shoreline of the lake.

To conclude, based on the analysis of phytoplankton and periphyton, the ecological state of Lake Durowskie remains stable, and has the potential to be better. The restoration methods that have been applied are really working, but it is a long procedure to achieve the state we expect.

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ANNEX 1. Comparison of phytoplankton species composition in different investigated years in July in Lake Durowskie.

Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015
Cyanoprokaryota - cyanobacteria								
<i>Anabaena affinis</i> Lemm.							+	
<i>Anabaena flos-aquae</i> Brebisson		+				+		
<i>Aphanizomenon aphanizomenoides</i> (Forti) Hort. & Kom.	+				+			
<i>Aphanizomenon flos-aquae</i> (L.) Ralfs	+	+	+	+			+	+
<i>Aphanizomenon gracile</i> Lemmerman	+							
<i>Aphanizomenon isatschenkoi</i> (Usacc.) Pros. - Lavrenko	+	+	+					
<i>Aphanocapsa grevillei</i> (Ber.) Rabenhorst		+						
<i>Aphanocapsa incerta</i> (Lemm.) Cronberg et Komarek	+	+	+			+		
<i>Arthrosphaera massartii</i> Kuff.		+						
<i>Chroococcus limneticus</i> Lemm.	+	+		+				
<i>Chroococcus turgidus</i> (Kütz.) Naeg.		+		+			+	+
<i>Cyanogranis feruginea</i> (Wawrik) Hind.		+	+					
<i>Gloeocapsa minuta</i> Lemm.							+	
<i>Jaaginema pseudogeminatum</i> (Schmid) Anagn. et Kom.			+	+				
<i>Limnothrix lauterbornii</i> (Schmidle) Anagn.		+						
<i>Limnothrix redekei</i> (Van Goor) Meffert	+		+	+	+	+	+	+
<i>Lyngbya hieronymusii</i> Lemm.		+						+
<i>Microcystis aeruginosa</i> Kützing	+			+	+	+	+	+
<i>Microcystis flos-aquae</i> (Wittrock) Kirchner			+			+		
<i>Jaaginema gracile</i> (Bocher) Anagn. et kom.		+						+
<i>Phormidium granulatum</i> Gardn. Anagn.	+	+	+		+			
<i>Phormidium tenue</i> (Agards ex Gomont) Anagn. et kom.		+						
<i>Planktolyngbya limnetica</i> (Lemm.) Kom. – Legn. Et Cronenberg		+	+	+	+	+	+	+
<i>Planktothrix agardhii</i> (D.C. ex Gom.) Anagn. et Kom.	+	+	+	+	+	+	+	+
<i>Pseudanabaena limnetica</i> (Lemm.) Kom.	+	+	+				+	+

Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen	+	+	+	+	+	+	+	+
<i>Gomphonema acuminatum</i> Ehr.					+	+	+	
<i>Gomphonema olivaceum</i> (Horn.) Breb.			+	+			+	
<i>Gomphonema parvulum</i> (Kütz.) Kütz.				+				
<i>Melosira varians</i> Ag.	+							
<i>Hippodonta capitata</i> (Ehr.) L-B, Metz. et Witk.						+		+
<i>Navicula cincta</i> (Ehr.) Ralfs	+	+	+			+	+	+
<i>Navicula mensiculus</i> Schumann	+							
<i>Navicula radiosa</i> Kützing			+	+	+	+	+	
<i>Naviula tripunctata</i> (O.F. Muller) Bory de Sain. Van.			+			+		+
<i>Nitzschia palea</i> (Kütz.) W. Smith				+	+	+	+	+
<i>Nitzschia recta</i> Hantzsch ex Rabenh.						+		
<i>Nitzschia sigmoidea</i> (Ehr.) W. Smith				+			+	+
<i>Nitzschia sinuata</i> (W. Sm.) Grunow				+				
<i>Pinnularia viridis</i> (Nitzsch) Ehr.			+				+	+
<i>Placoneis gastrum</i> (Ehr.) Meresch.		+						
<i>Rhopalodia gibba</i> (Ehr.) Muller						+		+
<i>Staurosira construens</i> Ehr.	+							
<i>Chlorophyta- green algae</i>								
<i>Ankistrodesmus falcatus</i> (Corda) Ralfs		+						
<i>Botryococcus braunii</i> Kütz.						+	+	+
<i>Characium aqungustatum</i> A. Braun		+		+	+	+	+	+
<i>Chlamydomonas globosa</i> Snow	+	+	+	+		+		
<i>Chlamydomonas passiva</i> Skuja			+			+	+	+
<i>Chlamydomonas reinhardtii</i> Dangeard		+						+
<i>Closterium acutum</i> var. <i>variabile</i> (Lemm.) Krieg.	+		+	+				+
<i>Coelasrum astroideum</i> De Notaris			+	+	+	+		+
<i>Coelastrum microporum</i> Naegel.			+					

Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015
<i>Coelastrum reticulatum</i> (Dang.) Senn	+	+					+	
<i>Cosmarium abbreviatum</i> Raciborski	+		+	+	+	+	+	
<i>Cosmarium exiguum</i> W. Archer		+						
<i>Cosmarium formosulm</i> Lund								+
<i>Cosmarium trilobulatum</i> Reinh								+
<i>Cosmarium margaritatum</i> (Turp.) Ralfs				+				
<i>Cosmarium phaseolus</i> Brebisson in Ralfs	+		+	+		+	+	+
<i>Cosmarium laeve</i> Rabenhorst					+			
<i>Cosmarium regnelli</i> Wille	+	+	+		+		+	+
<i>Crucigeniella rectnagulrais</i> (Naeg.) Kom.						+		
<i>Crucigenia tetrapedia</i> (Kirchner) W. et G.S. West			+					
<i>Desmodesmus communis</i> (Hegew.) Hegew.	+	+	+	+	+	+	+	+
<i>Desmodesmus grahamisii</i> (Heyning) Fott				+				
<i>Desmodesmus naegellii</i> (Meyen) Hegew.				+				
<i>Desmodesmus opoliensis</i> (Rchter) Hegew.			+			+		
<i>Desmodesmus subspicatus</i> (Chod.) Hegew. et Schmidt	+		+			+	+	+
<i>Dictyosphaerium pulchellum</i> Wood	+	+	+	+				
<i>Didymocystis plantonica</i> Korsikov				+				
<i>Elkatothrix gelatinosa</i> Wille			+	+		+	+	+
<i>Franceia ovais</i> (France) Lemm.							+	
<i>Golenkinia radiata</i> Chodat	+		+	+	+	+	+	+
<i>Kirchneriella contorta</i> var. <i>elegans</i> (Schmidle) Bohlin	+					+	+	+
<i>Koliella longiseta</i> (Vischer) Hindak	+							
<i>Lagerheimia ciliata</i> (Lag.) Chodat							+	
<i>Micractinium crassisetum</i> Hortobagyi				+				
<i>Micractinium pusillum</i> Fresenius				+				
<i>Mougeotia</i> sp.				+	+			
<i>Monoraphidium arcuatum</i> (Kors.) Hindak	+							

Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015
<i>Monoraphidium circinale</i> (Nyg.) Nygaard	+							
<i>Monoraphidium contortum</i> (Thur.) Kom.-Legn.	+	+	+	+	+	+	+	+
<i>Monoraphidium griffithii</i> (Berk.) Kom.-Legn.	+		+				+	+
<i>Monoraphidium irregulare</i> (G.M. Sm.) Kom.-Legn.	+		+					
<i>Monoraphidium komarkovae</i> Nygaard	+	+	+				+	+
<i>Monoraphidium minutom</i> (Nageli) Kom. - Legn.		+						
<i>Monoraphidium obtusum</i> (Kors.)Kom. - Legn.	+							
<i>Nephrocytium limneticum</i> (G. M. Sm.) G. M. Sm.				+				
<i>Oocystis lacustris</i> Chodat	+	+	+	+	+		+	+
<i>Oedogonium</i> sp.								+
<i>Palmelochette tenerrima</i> Kors.				+				
<i>Pandorina morum</i> (O.F. Müller) Bory			+			+		
<i>Pediastrum boryanum</i> (Turpin) Meneg.			+	+	+	+	+	+
<i>Pediastrum simplex</i> Meyen						+		
<i>Pediastrum duplex</i> Meyen						+	+	+
<i>Pediastrum tetras</i> (Ehr.) Ralfs			+				+	
<i>Phacotus lendneri</i> Chodat.				+				
<i>Phacotus lenticularis</i> (Ehr.) Stein	+			+	+	+	+	+
<i>Provasoliella saccata</i> (Skuja) Ettl					+			
<i>Provasiorella</i> sp.							+	
<i>Pteromonas angulosa</i> (Carter) Lemm.		+	+					
<i>Pteromonas cordiformis</i> Lemm.			+					
<i>Scenedesmus acuminatus</i> (Lager.) Chodat			+		+		+	+
<i>Scenedesmus bicaudatus</i> Dedusenko			+	+	+			
<i>Scenedesmus dimorphus</i> (Turp.) Kütz.		+		+				
<i>Scenedesmus ecornis</i> (Ehr.) Chod.			+	+	+		+	
<i>Scenedesmus obtusus</i> Meyen				+				
<i>Scenedesmus regularis</i> Swirensko		+						

Phytoplankton taxa	2008	2009	2010	2011	2012	2013	2014	2015
<i>Colacium vesiculosum</i> Ehr.		+		+		+		+
<i>Euglena caudata</i> Hübner					+			
<i>Euglena pisciformis</i> Klebs		+		+				+
<i>Phacus caudatus</i> Hubner								+
<i>Phacus pusillus</i> Lemm								+
<i>Phacus orbicularis</i> Hubner	+	+						
<i>Trachelomonas hispida</i> (Perty) Stein	+		+	+	+	+	+	+
<i>Trachelomonas planctonica</i> Swirensko	+		+	+				
<i>Trachelomonas volocina</i> Ehrenberg	+		+	+	+	+	+	
<i>Chrysophyceae - chrysophyces</i>								
<i>Chrysococcus rufescens</i> Klebs				+				
<i>Dinobryon bavaricum</i> Imhoff		+	+	+	+	+	+	
<i>Dinobryon crenulatum</i> W. et G.S. West		+	+	+				
<i>Dinobryon divergens</i> Imhof		+	+		+	+	+	+
<i>Dinobryon sociale</i> Ehrenberg	+		+	+		+		
<i>Erkenia subaequiciliata</i> Skuja	+	+	+		+	+	+	+

ANNEX 2. List of phytoplankton species from different taxonomical algal groups and their frequency in Lake Durowskie from 29th June to 3rd July 2015 (Inf – inflow; A2 – Aerator 2; Mid. 2 – Middle 2; Mid.1 – Middle 1; Outf – outflow; B1 – Beach 1; B2 – Beach 2; F – frequency).

	Site	Inf	A2	M2	M1	A1	Outf	B1	B2	F
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen		+		+	+			+		25
<i>Hippodonta capitata</i> (Ehr.) L-Bert., Metz. & Witkowski		+								6
<i>Navicula cincta</i> (Ehr.) Ralfs		+								6
<i>Naviula tripunctata</i> (Muller) Bory		+								6
<i>Nitzschia palea</i> (Kütz.) W. Smith		+								6
<i>Nitzschia sigmaoidea</i> (Ehr.) W. Smith		+								6
<i>Pinnularia viridis</i> (Nitzsch) Ehr.		+								6
<i>Rhopalodia gibba</i> (Ehr.) Muller		+								6
Chlorophyta - green algae										
<i>Botryococcus braunii</i> Kutz			+	+	+					31
<i>Characium aqungustum</i> A. Braun					+					6
<i>Chlamydomonas passiva</i> Skuja		+								6
<i>Chlamydomonas reinhardtii</i> Dangeard				+						13
<i>Closterium acutum</i> var. <i>variabile</i> (Lemm.) Krieg.			+			+		+		25
<i>Coelastrum astroideum</i> De Notaris		+	+	+	+	+				50
<i>Cosmarium phaseolus</i> Brebisson in Ralfs			+	+	+					19
<i>Cosmarium regnelli</i> Wille				+	+					25
<i>Cosmarium formosulum</i> Lund					+					13
<i>Cosmarium trilobulatum</i> Reinsch			+	+	+	+		+		31
<i>Desmodesmus communis</i> (Hegew.) Hegew.		+	+	+	+	+	+		+	63
<i>Desmodesmus subspicatus</i> (Chod.) Hegew. et Schmidt					+			+		13
<i>Elkatothrix gelatinosa</i> Wille		+	+	+	+	+		+	+	81
<i>Golenkinia radiata</i> Chodat		+	+		+	+			+	38
<i>Kirchneriella contorta</i> var. <i>elegans</i> (Schmidle) Bohlin						+		+		13
<i>Monoraphidium contortum</i> (Thur.) Kom.-Legn.		+	+	+		+			+	31
<i>Monoraphidium griffithii</i> (Berk.) Kom.-Legn.						+				6
<i>Monoraphidium komarkovae</i> Nygaard						+				6
<i>Oocystis lacustris</i> Chodat			+	+	+	+			+	44

	Site	Inf	A2	M2	M1	A1	Outf	B1	B2	F
<i>Oedogonium sp.</i>				+				+		13
<i>Pediastrum boryanum</i> (Turpin) Meneg.	+			+	+	+	+			50
<i>Pediastrum duplex</i> Meyen					+	+		+		19
<i>Phacotus lenticularis</i> (Ehr.) Stein	+	+	+	+	+	+	+		+	88
<i>Scenedesmus acuminatus</i> (Lager.) Chodat	+	+						+		19
<i>Sphaerocystis planctonica</i> (Korsikov) Bourrelly			+	+	+	+	+		+	81
<i>Tetraedron caudatum</i> (Corda) Hansgirg				+		+				13
<i>Tetraedron minimum</i> (A. Br.) Hansgirg	+	+	+	+	+	+				38
<i>Tetrastrum staurogeanieforme</i> (Schroed.) Lemm.				+						13
<i>Staurastrum gracile</i> Ralfs	+	+	+	+	+	+				50
<i>Cryptophyta - cryptophytes</i>										
<i>Cryptomonas erosa</i> Ehrenberg	+	+	+	+	+	+	+	+	+	100
<i>Cryptomonas marssonii</i> Skuja	+	+	+			+	+	+	+	44
<i>Cryptomonas ovata</i> Ehrenberg								+		6
<i>Cryptomonas rostrata</i> Troitzkaja emend I. Kiselev	+	+	+	+	+	+	+	+	+	94
<i>Rhodomonas minuta</i> Skuja	+	+	+	+	+	+	+	+	+	88
<i>Dinophyta - dinoflagellates</i>										
<i>Peridinopsis berolinense</i> (Lemm.) Bourrelly			+	+			+		+	25
<i>Ceratium hirundinella</i> (F. B. Müller) Bergh	+	+	+	+	+	+	+	+		94
<i>Ceratium cornutum</i> (Ehr.) Clap. & Lachman			+	+	+					81
<i>Gymnodinium aeruginosum</i> Stein	+									6
<i>Peridiniopsis cuningtonii</i> Lemm.	+	+	+	+	+	+	+	+	+	100
<i>Peridinium cinctum</i> (O.F. Müller) Ehrenberg	+	+	+	+	+	+	+	+	+	94
<i>Peridinopsis elpatiewskyi</i> (Ostenf.) Bourrelly	+	+	+	+	+	+	+	+	+	100
<i>Euglenophyta - euglenoids</i>										
<i>Trachelomonas hispida</i> (Perty) Stein	+	+	+				+	+	+	44
<i>Euglena pisciformis</i> Klebs	+	+		+	+					25
<i>Phacus caudatus</i> Hubner			+							6

	Site	Inf	A2	M2	M1	A1	Outf	B1	B2	F
<i>Phacus pusillus</i> Lemm.				+				+		13
<i>Colacium vesiculosum</i> Ehr.				+	+				+	19
<i>Chrysophyceae - chrysophyces</i>										
<i>Erkenia subaequiciliata</i> Skuja		+	+	+	+	+	+	+	+	88
<i>Dinobryon divergens</i> Imhof		+	+	+	+	+	+	+		63

ANNEX 3. Average number of phytoplankton species cells (ind./L) from different depth in Lake Durowskie from 29th June to 03rd July 2015 (Inf – inflow; A2 – Aerator 2; Mid. 2 – Middle 2; Mid.1 – Middle 1; Outf – outflow; B1 – Beach 1; B2 – Beach 2).

	Depth	0-2m	0-2m	0-2m	0-2m	0-2m	0-2m	0-2m	0-2m
	Site	Inf	A2	Mid.2	Mid.1	A1	Outf	B1	B2
<i>Cyanoprokaryota - cyanobacteria</i>									
<i>Aphanizomenon flos-aquae</i> (L.) Ralfs		191200	12000	1600	1067	800	800	.	1600
<i>Chroococcus turgidus</i> (Kütz.) Naeg.		.	.	.	267
<i>Limnothrix redekelei</i> (Van Goor) Meffert		1714400	37067	82133	20267	82933	17600	20800	29600
<i>Lyngbya hieronymusii</i> Lemm.	
<i>Microcystis aeruginosa</i> Kützing		800
<i>Planktolyngbya limnetica</i> (Lemm.) Kom. – Legn. Et Cronenberg		3200	.
<i>Planktothrix agardhii</i> (D.C. ex Gom.) Anagn. et Kom.		92800	4533	12000	3733	5867	2400	3200	3200
<i>Pseudanabaena limnetica</i> (Lemm.) Kom.		276000
<i>Woronichina naegeliana</i> (Unger) Elenkin		.	.	267
<i>Jaaginema gracilis</i> (Bocher) Anagn. et Kom.		267	.	.	.
Total		2275200	53600	96000	25333	89867	20800	27200	34400
<i>Bacillariophyce - diatoms</i>									
<i>Amphora ovalis</i> Kützing		800
<i>Amphora copulata</i> (Kutz.) Schoeman & Archibald		1600	800
<i>Asterionella formosa</i> Hasall		.	267	.	800	1333	.	.	.
<i>Caloneis amphisbaena</i> (Bory) Cleve		800

	Site	Inf	A2	Mid.2	Mid.1	A1	Outf	B1	B2
<i>Cocconeis placentula</i> Ehr.		800
<i>Cyclotella ocellata</i> Pant.		4800	533	1867	1067	533	.	800	800
<i>Cyclotella radiososa</i> (Grun.) Lemm.		20800	4533	7467	3200	7200	11200	1600	5600
<i>Cymbella minuta</i> Hilse ex Rabenhorst		.	.	267	267
<i>Fragilaria crotonensis</i> Kitton		48000	4533	6667	8800	31467	14400	20800	12000
<i>Fragilaria pinnata</i> Ehr.		.	.	.	267
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot		9600	24000	28000	32000	42133	.	28800	42400
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen		800	.	533	267	.	.	28000	.
<i>Hippodonta capitata</i> (Ehr.) L-Bert., Metz. & Witkowski		800
<i>Navicula cincta</i> (Ehr.) Ralfs		800
<i>Naviula tripunctata</i> (Muller) Bory		800
<i>Nitzschia palea</i> (Kütz.) W. Smith		2400
<i>Nitzschia sigmoidea</i> (Ehr.) W. Smith		800
<i>Pinnularia viridis</i> (Nitzsch) Ehr.		800
<i>Rhopalodia gibba</i> (Ehr.) Muller		800
	Total	93600	33867	44800	46667	82667	25600	81600	61600
<i>Chlorophyta - green algae</i>									
<i>Botryococcus braunii</i> Kutzing		.	1600	1867	1067
<i>Characium aqngustatum</i> A. Braun		.	.	.	267
<i>Chlamydomonas passiva</i> Skuja		800
<i>Chlamydomonas reinhardtii</i> Dangeard		.	.	4000
<i>Closterium acutum</i> var. <i>variabile</i> (Lemm.) Krieg.		.	533	.	.	267	.	.	.
<i>Coelastrum astroideum</i> De Notaris		800	533	1067	533	533	.	1600	.
<i>Cosmarium phaseolus</i> Brebisson in Ralfs		.	267	267	267
<i>Cosmarium regnelli</i> Wille		.	.	800	533
<i>Cosmarium formosulum</i> Lund		.	.	533
<i>cosmarium trilobulatum</i> Reinsch		.	533	267	267	533	.	.	.
<i>Desmodesmus communis</i> (Hegew.) Hegew.		1600	533	800	267	533	1600	800	800

	Site	Inf	A2	Mid.2	Mid.1	A1	Outf	B1	B2
<i>Peridinopsis berolinense</i> (Lemm.) Bourrelly	.	267	267	.	1333	.	.	.	2400
<i>Ceratium hirundinella</i> (F. B. Müller) Bergh	1600	4533	8533	4000	4800	4000	4000	.	.
<i>Ceratium cornutum</i> (Ehr.) Clap. & Lachman	.	800	800	800	800	.	800	.	.
<i>Gymnodinium aeruginosum</i> Stein	800
<i>Peridiniopsis cuningtonii</i> Lemm.	8000	38133	40533	20267	68800	64800	12000	20000	
<i>Peridinium cinctum</i> (O.F. Müller) Ehrenberg	4800	20267	23733	14133	52533	80000	6400	4000	
<i>Peridinopsis elpatiewskyi</i> (Ostenf.) Bourrelly	4800	3200	3467	4267	12000	36000	12800	13600	
Total	20000	67200	77333	43467	140267	184800	36000	40000	
Euglenophyta - euglenoids									
<i>Trachelomonas hispida</i> (Perty) Stein	1600	533	1800	.	.	800	800	800	
<i>Euglena pisciformis</i> Klebs	800	533	.	267	800
<i>Phacus caudatus</i> Hubner	.	.	2400
<i>Phacus pusillus</i> Lemm.	.	.	800	.	.	.	800	.	.
<i>Colacium vesiculosum</i> Ehr.	.	.	1867	267	2400
Total	2400	1067	6867	533	800	800	1600	3200	
Chrysophyceae - chrysophyces									
<i>Erkenia subaequiciliata</i> Skuja	150400	17333	15467	12000	19467	4000	79200	4200	
<i>Dinobryon divergens</i> Imhof	15200	1067	267	533	1600	4000	1600	.	.
Total	165600	18400	15733	12533	21067	8000	80800	4200	

**ANNEX 4. Average biomass of phytoplankton species (mg/L) from different depth in Lake Durowskie from 29th June to 03rd July 2015
 (Inf – inflow; A2 – Aerator 2; Mid. 2 – Middle 2; Mid.1 – Middle 1; Outf – outflow; B1 – Beach 1; B2 – Beach 2).**

Depth	0-2m							
	Site	Inf	A2	Mid.2	Mid.1	A1	Outf	B1
Cyanoprokaryota - cyanobacteria								
<i>Aphanizomenon flos-aquae</i> (L.) Ralfs	0.375	0.024	0.003	0.002	0.002	0.002	.	0.003
<i>Chroococcus turgidus</i> (Kütz.) Naeg.	.	.	.	0.002
<i>Limnothrix redekelei</i> (Van Goor) Meffert	.	.	.	0.0003
<i>Lyngbya hieronymusii</i> Lemm.	0.538	0.012	0.025	0.006	0.026	0.006	0.007	0.009
<i>Microcystis aeruginosa</i> Kützing	0.001
<i>Planktolyngbya limnetica</i> (Lemm.) Kom. – Legn. Et Cronenberg	0.001	.
<i>Planktothrix agardhii</i> (D.C. ex Gom.) Anagn. et Kom.	0.117	0.006	0.017	0.005	0.007	0.003	0.004	0.004
<i>Pseudanabaena limnetica</i> (Lemm.) Kom.	0.087
<i>Woronichina naegeliana</i> (Unger) Elenkin	.	.	0.0003
<i>Jaaginema gracilis</i> (Bocher) Anagn. et Kom.	0.001	.	.	.
Total	0.204	0.006	0.017	0.005	0.008	0.003	0.012	0.004
Bacillariophycee - diatoms								
<i>Amphora ovalis</i> Kützing	0.001
<i>Amphora copulata</i> (Kutz.) Schoeman & Archibald	0.0002	0.0008
<i>Asterionella formosa</i> Hasall	.	0.00003	.	0.0001	0.0002	.	.	.
<i>Caloneis amphisbaena</i> (Bory) Cleve	0.009
<i>Cocconeis placentula</i> Ehr.	0.001
<i>Cyclotella ocellata</i> Pant.	0.005	0.001	0.011	0.001	0.001	.	0.0009	0.0008
<i>Cyclotella radiososa</i> (Grun.) Lemm.	0.026	0.006	0.011	0.004	0.009	0.014	0.002	0.007
<i>Cymbella minuta</i> Hilse ex Rabenhorst	.	.	0.0003	0.0003
<i>Fragilaria crotonensis</i> Kitton	0.022	0.002	0.004	0.004	0.014	0.007	0.009	0.005
<i>Fragilaria pinnata</i> Ehr.	.	.	.	0.0003
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot	0.013	0.232	0.041	0.044	0.057	0.038	0.039	0.058

	Site	Inf	A2	Mid.2	Mid.1	A1	Outf	B1	B2
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen		0.002	.	0.003	0.001
<i>Hippodonta capitata</i> (Ehr.) L-Bert., Metz. & Witkowski		0.002
<i>Navicula cincta</i> (Ehr.) Ralfs		0.01
<i>Naviula tripunctata</i> (Muller) Bory		0.012
<i>Nitzschia palea</i> (Kütz.) W. Smith		0.03
<i>Nitzschia sigmoidea</i> (Ehr.) W. Smith		0.013
<i>Pinnularia viridis</i> (Nitzsch) Ehr.		0.002
<i>Rhopalodia gibba</i> (Ehr.) Muller		0.001
	Total	0.149	0.240	0.070	0.055	0.081	0.059	0.0511	0.0716
<i>Chlorophyta - green algae</i>									
<i>Botryococcus braunii</i> Kutzin		.	0.007	0.007	0.002
<i>Characium aqngustatum</i> A. Braun		.	.	.	0.0002
<i>Chlamydomonas passiva</i> Skuja		0.001
<i>Chlamydomonas reinhardtii</i> Dangeard		.	.	0.002
<i>Closterium acutum</i> var. <i>variabile</i> (Lemm.) Krieg.		.	0.001
<i>Coelastrum astroideum</i> De Notaris		0.003	0.002	0.004	0.002	0.002	.	0.005	.
<i>Cosmarium phaseolus</i> Brebisson in Ralfs		.	0.001	0.001	0.001
<i>Cosmarium regnelli</i> Wille		.	.	0.001	0.001
<i>Cosmarium formosulm</i> Lund		.	.	0.007
<i>cosmarium trilobulatum</i> Reinsch		.	0.0003	0.001	0.0003	0.003	.	.	.
<i>Desmodesmus communis</i> (Hegew.) Hegew.		0.001	0.001	0.001	0.0003	.	0.001	0.0007	0.001
<i>Desmodesmus subspicatus</i> (Chod.) Hegew. et Schmidt		.	.	.	0.0003
<i>Elkatothrix gelatinosa</i> Wille		0.005	0.004	0.003	0.002	0.002	.	0.004	0.001
<i>Golenkinia radiata</i> Chodat		0.006	0.001	.	0.001	0.0004	.	0.002	0.001
<i>Kirchneriella contorta</i> var. <i>elegans</i> (Schmidle) Bohlin		.	.	.	0.00003
<i>Monoraphidium contortum</i> (Thur.) Kom.-Legn.		0.001	0.0003	.	.	0.00003	.	0.0003	0.0001
<i>Monoraphidium griffithii</i> (Berk.) Kom.-Legn.		0.001	.	.	.
<i>Monoraphidium komarkovae</i> Nygaard		0.002	.	.	.

	Site	Inf	A2	Mid.2	Mid.1	A1	Outf	B1	B2
<i>Oocystis lacustris</i> Chodat	.	0.001	0.001	0.001	0.009	.	.	.	0.045
<i>Oedogonium</i> sp.	.	.	0.013
<i>Pediastrum boryanum</i> (Turpin) Meneg.	0.024	.	0.080	0.024	0.118	0.024	0.024	.	.
<i>Pediastrum duplex</i> Meyen	0.008
<i>Phacotus lenticularis</i> (Ehr.) Stein	0.024	0.012	0.001	0.002	0.003	0.001	0.003	0.003	.
<i>Scenedesmus acuminatus</i> (Lager.) Chodat	0.001
<i>Sphaerocystis planctonica</i> (Korsikov) Bourrelly	.	0.021	0.015	0.016	0.057	0.071	0.002	0.029	.
<i>Tetraedron caudatum</i> (Corda) Hansgirg	.	.	0.0000 3	.	0.00003
<i>Tetraedron minimum</i> (A. Br.) Hansgirg	0.0001	.	0.0000 3	0.0002	0.00003
<i>Tetrastrum staurogeanieforme</i> (Schroed.) Lemm.	.	.	0.0001
<i>Staurastrum gracile</i> Ralfs	0.002	0.006	0.003	0.001	0.021
Total	0.0681	0.056	0.140	0.053	0.226	0.097	0.041	0.0801	
Cryptophyta - cryptophytes									
<i>Cryptomonas erosa</i> Ehrenberg	0.036	0.210	0.097	0.163	0.238	0.071	0.11	0.141	.
<i>Cryptomonas marssonii</i> Skuja	0.033	0.001	.	.	0.001	0.009	0.001	0.005	.
<i>Cryptomonas ovata</i> Ehrenberg	0.002	.	.
<i>Cryptomonas rostrata</i> Troitzkaja emend I. Kiselev	0.02	0.035	0.032	0.007	0.026	0.012	0.007	0.007	.
<i>Rhodomonas minuta</i> Skuja	0.002	0.001	0.019	0.001	0.001	0.0006	0.003	0.004	.
Total	0.091	0.247	0.148	0.171	0.266	0.0926	0.123	0.157	
Dinophyta - dinophytes									
<i>Peridinopsis berolinense</i> (Lemm.) Bourrelly	.	0.002	0.003	.	0.001	.	0.037	0.022	.
<i>Ceratium hirundinella</i> (F. B. Müller) Bergh	.	0.181	0.822	0.310	0.464	0.387	0.077	.	.
<i>Ceratium cornutum</i> (Ehr.) Clap. & Lachman	0.155	0.306	0.072	0.125	0.072	0.072	.	.	.
<i>Gymnodinium aeruginosum</i> Stein	0.001
<i>Peridiniopsis cuningtonii</i> Lemm.	0.092	0.441	0.467	0.234	0.796	0.75	0.139	0.231	.
<i>Peridinium cinctum</i> (O.F. Müller) Ehrenberg	0.127	0.538	0.628	0.376	1.394	2.123	0.17	0.106	.
<i>Peridinopsis elpatiewskyi</i> (Ostenf.) Bourrelly	0.038	0.102	0.026	0.034	0.096	0.288	0.102	0.109	.

	Total	0.413	1.570	2.019	1.080	2.823	3.62	0.525	0.468
Euglenophyta - euglenoids									
<i>Trachelomonas hispida</i> (Perty) Stein	0.001	0.001	0.002	.	.	0.0008	0.0008	0.001	
<i>Euglena pisciformis</i> Klebs	0.001	.	.	0.0003	0.0005
<i>Phacus caudatus</i> Hubner	.	.	0.002	.	.	.	0.002	.	.
<i>Phacus pusillus</i> Lemm.	.	.	0.001
<i>Colacium vesiculosum</i> Ehr.	.	.	0.003	0.0002	.	.	.	0.002	
Total	0.002	0.001	0.008	0.001	0.0005	0.0008	0.0028	0.003	
Chrysophyceae - chrysophyces									
<i>Erkenia subaequiciliata</i> Skuja	0.008	0.001	0.001	0.001	0.001	0.0002	0.004	0.0002	
<i>Dinobryon divergens</i> Imhof	0.007	0.0005	0.0001	0.0003	0.001	0.002	0.0007	.	.
Total	0.015	0.002	0.001	0.001	0.002	0.0022	0.0047	0.0002	

ANNEX 5. Comparison of periphyton species composition in different investigated sites from 29th June to 4th July 2015 in Lake Durowskie.

Diatom taxa	Site 1	Site 2	Site 3	Site 4	Site 5	Site 6	Site 7	Site 8	Site 9	Site 10	pH	O	T
<i>Achnanthes conspicua</i> Mayer	3	2	7
<i>Achnanthes exigua</i> Grun.	.	+	+	+	+	+	+	+	+	+	4	1	7
<i>Achnanthes hungarica</i> (Grun.) Grun.	.	.	.	+	.	.	+	.	.	.	4	4	6
<i>Achnanthes lanceolata</i> (Breb.) Grun.	+	+	+	+	.	+	+	+	.	.	4	3	5
<i>Achnanthes lanceolata v. elliptica</i> Cleve sensu Straub	.	+	.	.	.	+	4	-	-
<i>Achnanthes minutissima</i> Kütz.	+	+	+	+	+	+	+	+	+	+	3	1	7
<i>Achnanthes minutissima</i> var. <i>affinis</i> (Grun.) Lange-Bertalot	+	+	+	.	.	4	-	-
<i>Achnanthes minutissima</i> var. <i>gracillina</i> (Meister) Lange-Bertalot	.	.	.	+	.	+	.	+	.	.	4	-	1
<i>Amphipleura pellucida</i> (Kuetzing) Kuetzing	4	2	2
<i>Amphora ovalis</i> Kütz.	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Amphora pediculus</i> (Kütz.) Grun.	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Amphora pellucida</i> Kütz.	.	+	-	-	-
<i>Asterionella formosa</i> Hass	+	.	.	.	+	.	.	+	+	+	4	2	5

Diatom taxa	Site 1	Site 2	Site 3	Site 4	Site 5	Site 6	Site 7	Site 8	Site 9	Site 10	pH	O	T
<i>Aulacoseira granulata</i> (Ehr.) Ralfs	.	+	4	3	5
<i>Aulcoseira granulata</i> var. <i>angustissima f.curvata</i> (O. Mül.) Sim.	4	3	5
<i>Caloneis bacillum</i> (Grun.) Meresz.	.	+	+	+	.	+	4	2	4
<i>Caloneis silicula</i> (Ehr.) Cleve	.	+	.	+	4	2	4
<i>Coccconeis euglypta</i> (Ehr.) Clevei	+	.	.	-	-	-
<i>Coccconeis pediculus</i> Ehr	.	+	.	+	+	+	+	.	+	+	4	2	5
<i>Coccconeis placentula</i> Ehr.	+	+	+	+	+	+	+	+	+	+	4	3	5
<i>Coccconeis placentula</i> var. <i>linearis</i> Ehr.	.	.	+	-	-	-
<i>Coccconeis placentula</i> var. <i>lineata</i> (Ehr.)	.	+	+	.	.	+	.	+	.	.	4	3	5
<i>Coccconeis placentula</i> var. <i>pseudolineata</i> Geitler	+	+	.	.	.	+	+	.	.	.	-	-	-
<i>Cyclotella meneghiniana</i> Kütz.	.	.	+	+	+	+	+	+	+	+	4	5	5
<i>Cyclotella ocellata</i> Pant.	+	+	+	+	+	+	+	+	+	.	4	1	4
<i>Cyclotella operculata</i> (Ag.) Kütz.	.	+	+	+	.	.	+	+	+	+	-	-	-
<i>Cyclotella radiososa</i> (Grun.) Lemm.	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Cyclotella stelligera</i> Cl. et Grun.	+	.	.	0	-	-
<i>Cymatopleura solea</i> (Breb.) W. Smith	+	.	.	.	+	+	4	3	5
<i>Cymbella affinis</i> Kütz.	+	.	+	+	+	+	+	+	+	+	4	1	5
<i>Cymbella caespitosum</i> (Kütz.) Brun	.	.	.	+	0	-	7
<i>Cymbella cistula</i> (Her.) Kirchner	+	+	+	.	+	.	.	.	+	.	4	2	5
<i>Cymbella lanceolata</i> (Ehr.) Kirchner	+	.	.	.	+	.	.	.	+	+	4	1	7
<i>Cymbella microcephala</i> Grun.	+	+	+	.	.	+	+	+	.	.	4	1	4
<i>Cymbella minuta</i> Hilse	+	+	+	+	+	+	+	+	+	+	3	-	-
<i>Cymbella prostrata</i> (Berkeley) Cleve	+	.	4	1	5
<i>Cymbella turgida</i> (Greg.) Cleve	+	-	-	-
<i>Diatoma tenuis</i> Agardh	+	.	.	.	+	.	.	.	+	+	4	3	5
<i>Diatoma vulgaris</i> Bory	.	+	.	+	+	+	+	+	+	+	5	2	4
<i>Diatoma vulgaris</i> Bory Morphotyp <i>ovalis</i>	-	-	-
<i>Diploneis elliptica</i> (Kütz.) Cl.	+	.	.	.	+	+	4	1	3
<i>Encynomena silesiacum</i> (Bleisch in Rabenh.) D. G. Mann	-	-	-
<i>Eunotia exigua</i> (Breb.) Rabenh.	+	.	.	.	+	.	.	.	+	+	1	2	7
<i>Eunotia faba</i> (Ehr.) Grun.	+	.	.	.	+	.	2	1	2
<i>Eunotia intermedia</i> (Krasske) Norpel I Lange-Bertalot	.	.	.	+	2	-	1
<i>Eunotia lunaris</i> Ehr.	+	-	-	-

Diatom taxa	Site 1	Site 2	Site 3	Site 4	Site 5	Site 6	Site 7	Site 8	Site 9	Site 10	pH	O	T
<i>Eunotia praerupta</i> Ehr.	+	+	+	+	+	+	+	+	+	+	2	1	2
<i>Eunotia tenella</i> (Grun.) Hustedt	.	.	.	+	2	1	1
<i>Fragilaria capitata</i> Ehr.	.	.	+	-	-	-	-
<i>Fragilaria capucina</i> (Desm.) Rabenhorst	+	+	+	+	+	+	+	.	+	3	-	3	
<i>Fragilaria capucina</i> v. <i>amphcephala</i>	+	-	-	-	-
<i>Fragilaria constricta</i> Ehr.	+	.	.	2	-	1	
<i>Fragilaria construens</i> (Ehr.) Grun.	.	.	+	4	1	4	
<i>Fragilaria crotonensis</i> Kitton	+	+	+	+	+	+	+	+	+	4	2	3	
<i>Fragilaria exigua</i> Grun.	+	.	3	1	1	
<i>Fragilaria martyi</i> (Heribaud) Lange-Bertalot	.	+	+	-	-	-	-
<i>Fragilaria pinnata</i> Ehr.	+	+	+	+	+	+	+	+	.	+	4	1	7
<i>Fragilaria ulna</i> (Nitzsch) Lange-Bertalot	+	+	+	+	+	+	+	+	.	+	4	3	7
<i>Fragilaria ulna</i> var. <i>angustissima</i> Sippen	+	+	+	.	.	+	.	+	.	4	2	7	
<i>Fragilaria ulna</i> var. <i>biceps</i> (Nitzsch) Lange-Bertalot	+	-	-	-	-
<i>Fragilaria vaucheriae</i> (Kütz.) Carlson	-	-	-	-
<i>Gomphonema acuminatum</i> Ehr.	+	.	.	+	+	+	.	+	.	+	4	2	5
<i>Gomphonema angustatum</i> (Kütz.) Rabenhorst	.	.	.	+	-	-	-	-
<i>Gomphonema angustum</i> (Kütz.) Rabenhorst	+	4	1	1	
<i>Gomphonema augur</i> Ehr.	+	.	.	+	4	1	4	
<i>Gomphonema gracile</i> Ehr.	+	+	.	.	+	.	.	.	+	3	1	3	
<i>Gomphonema intricatum</i> Kütz.	.	.	.	+	-	-	-	-
<i>Gomphonema italicum</i> Kütz.	-	-	-	-
<i>Gomphonema micropus</i> Kütz.	+	.	.	.	+	4	2	5	
<i>Gomphonema olivaceoides</i> Hustedt	+	+	.	+	.	+	.	.	+	+	-	-	-
<i>Gomphonema olivaceum</i> (Horn.) Breb.	+	+	+	+	+	.	+	+	+	5	2	5	
<i>Gomphonema parvulum</i> (Kütz.) Kütz.	+	+	.	+	+	.	+	+	+	3	4	5	
<i>Gomphonema truncatum</i> Ehr.	.	+	4	2	4	
<i>Hantzschia amphioxys</i> (Ehr.) Grun.	+	.	+	3	2	7	
<i>Hipodonta capitata</i> (Ehr.) Lange-Bertalot	-	-	-	-
<i>Mastogloia smithii</i> Twaites ex W. Smith	+	.	.	.	4	-	-	
<i>Meridion circulare</i> Ag.	.	+	+	+	.	+	+	.	.	4	2	7	
<i>Navicula agrestis</i> Hustedt	+	+	3	-	-	
<i>Navicula anglica</i> Ralfs	+	.	-	-	-	-

Diatom taxa	Site 1	Site 2	Site 3	Site 4	Site 5	Site 6	Site 7	Site 8	Site 9	Site 10	pH	O	T
<i>Navicula capitata</i> Patrick in Patrick & Reimer	.	+	+	.	.	+	+	+	+	+	4	3	4
<i>Navicula cincta</i> (Ehr.) Ralfs	+	.	+	.	+	.	.	+	+	+	4	3	5
<i>Navicula cryptocephala</i> Kütz.	+	.	.	.	+	.	.	+	+	+	3	3	7
<i>Navicula dicephala</i> (Ehr.) W. Sm.	+	-	-	-	-
<i>Navicula gastrum</i>	+	4	4	5
<i>Navicula gregaria</i> Donkin	+	.	+	.	.	4	4	5
<i>Navicula integra</i> (W. Smith) Ralfs	3	3	5
<i>Navicula placentula</i> (Placeneis)	+	+	4	2	5
<i>Navicula pseudoanglica</i> Lange-Bertalot	4	1	4
<i>Navicula radiosa</i> Kütz.	+	+	+	+	+	+	+	+	+	+	3	2	4
<i>Navicula reinhardtii</i> Grun.	.	.	+	.	.	+	.	.	.	+	5	2	5
<i>Navicula tenella</i> Brébisson ex Kützing	-	-	-	-
<i>Navicula tripunctata</i> (O. F. Müller) Bory	+	+	+	+	+	+	+	+	+	+	4	2	5
<i>Navicula viridula</i> (Kütz.) Ehr.	+	.	+	.	+	.	4	2	5
<i>Nitzschia acicularis</i> (Kütz.) W. Sm	+	+	+	4	4	5
<i>Nitzschia amphibia</i> Grun.	+	+	.	+	+	.	.	.	+	+	4	3	5
<i>Nitzschia incospicua</i> Grun.	+	+	.	.	+	4	3	5
<i>Nitzschia intermedia</i> Hantzsch	3	-	5
<i>Nitzschia micropus</i> Kütz.	+	-	-	-	-
<i>Nitzschia palea</i> (Kütz.) W. Sm.	+	.	+	+	.	.	+	+	.	.	3	4	6
<i>Nitzschia paleacea</i> Grun.	+	.	+	+	.	+	+	.	.	.	4	3	5
<i>Nitzschia recta</i> Hantzsch	+	.	.	.	+	+	+	+	+	.	4	2	7
<i>Nitzschia sigmoidea</i> (Ehr.) W. Sm.	.	.	+	+	+	.	+	.	+	+	4	3	5
<i>Pinnularia gibba</i> Ehr.	.	+	3	3	7
<i>Pinnularia maior</i> (Kütz.) Cleve	+	.	.	3	2	4
<i>Pinnularia viridis</i> (Nitzsch) Ehr.	.	+	+	+	.	+	+	.	.	.	3	3	7
<i>Placoneis placentula</i> (Ehr.) Cox	-	-	-
<i>Rhoicosphaenia abbreviata</i> (Ag.) Lange-Bertalot	.	+	.	+	4	2	5
<i>Rhoicosphaenia curvata</i> (Kütz.) Grun.	.	.	+	-	-	-	-
<i>Rhopalodia gibba</i> (Ehr.) Mueller	.	+	+	.	+	5	3	5
<i>Stauroneis phoenicentron</i> Ehr.	+	.	+	.	.	+	.	+	.	.	3	3	4
<i>Stephanodiscus astraea</i> (Ehrenberg) Grunow	+	+	.	+	.	-	-	-

Diatom taxa	Site 1	Site 2	Site 3	Site 4	Site 5	Site 6	Site 7	Site 8	Site 9	Site 10	pH	O	T
<i>Stephanodiscus hantzschii</i> Grun.	+	.	+	+	.	.	.	+	.	+	5	4	6
<i>Surirella minuta</i> Breb.	.	.	+	4	3	5
<i>Surirella ovalis</i> Breb.	+	.	.	.	+	.	4	4	5
<i>Syndera capitata</i> Ehrenberg	-	-	-
<i>Synedra ulva</i> (Nitzsch)	-	-	-
<i>Tabellaria fenestrata</i> (Lyngb.) Kütz.	+	+	.	.	.	+	.	+	.	.	3	1	2
<i>Tabellaria flocculosa</i> (Roth) Kütz.	+	2	1	3